

Association of Drinking Water and Enteric Fever: A Disguised Source of Infection

FATIMA TUZ ZAHRA, SIDRAH SALEEM, MUHAMMAD IMRAN, NIDA JAVED, AYESHA GHAZAL, SAADIA CH*

Department of Microbiology, University of Health Sciences, Lahore, 54000, Pakistan.

*Lahore Medical & Dental College, Lahore

Correspondence to Dr. Fatima Tuz Zahra, Email: fatimatuzzahra2022@gmail.com, Phone: +92 3014465892

ABSTRACT

Background: Enteric fever is an illness caused by *Salmonella enterica* serovars *Typhi* and *Paratyphi A* and *B*. The mode of transmission is the ingestion of contaminated food and water.

Aim: To check that whether drinking water is the source of infection or not.

Methodology: 202 water samples were collected from the various areas of Lahore. 120 samples were taken from houses and the neighbourhood of confirmed typhoid patients and 82 samples were randomly collected. The samples were centrifuged, enriched and then subcultured on XLD media. The isolated colonies were identified by biochemical reactions. The sero-typing confirmed *Salmonella* species were done.

Results: The results showed that 23% of the drinking water in Lahore was fit for drinking, 76% samples were contaminated. The organisms isolated were *Serratia* spp. 22%, *Enterobacter* spp. 22%, *E. coli* 16%, *Citrobacter* spp. 12%, *Pseudomonas* spp. 9%, *Proteus* spp. 6%, *Klebsiella* spp. 4%, *Salmonella Paratyphi A* and *Salmonella Rubislaw* as 1%. No *Salmonella Typhi* was isolated from any water sample.

Conclusion: The presences of large coliforms in drinking water were indicative of sewerage contamination. The *Salmonella Paratyphi A* might be source of infection in that specific area but overall the results suggested that drinking water of Lahore was not acting as a source of infection for Enteric caused by *Salmonella Typhi*.

Keywords: Water-borne, Infection, Enteric fever, coliforms.

INTRODUCTION

Salmonella species are a member of family 'Enterobacteriaceae'. *Salmonella Typhi* is an encapsulated Gram negative rod, non-spore forming and motile. *Salmonella enterica* and *Salmonella bongori* are the two species causing human infections¹. Within *Salmonella enterica* subspecies, serovars *Typhi* and *Paratyphi A* are the major cause of enteric fever (also known as typhoid fever).

Enteric fever is a significant public health issue in the developing and under developed countries². Worldwide, the incidence of *Salmonella Typhi* cases is about 21 million but the rate of morbidity and mortality is very high in the endemic regions i.e, Southeast Asia and Sub-saharan Africa³. The rate reported is around 80% in Asia and 20% in Africa and Latin America². Pakistan also has a high incidence of typhoid fever with significant levels of morbidity and mortality. Bacteremia in children is recorded as high as 1000 positive cases per 100,000 children in Karachi⁴.

An important vehicles of transmission is the natural waters⁵. *S. Typhi* can be acquired by drinking / swimming in contaminated water and eating food washed with contaminated water¹. Majority of outbreaks in the developed countries are due to the intake of untreated or inefficiently treated water. Reports have estimated that approximately 1.1 billion people use unsafe water worldwide. This unsafe water, sanitation and hygiene results in 88% of diarrheal disease in the world. In many areas, the spread of enteric fever is also as a result of ingestion of contaminated food caused by indirect contamination. This indirect contamination can occur while irrigation and even while washing the food with contaminated water⁵.

In 2017, an outbreak of XDR *S. Typhi* was reported in Sindh, Pakistan. After the higher incidence of XDR cases, the water supply of the neighborhood of the cases was screened for the presence of *S. Typhi*. Polymerase Chain Reaction of the water sample indicated water contamination which was acting as a reservoir for the outbreak⁴.

The purpose of this study was to identify the areas with high incidence of typhoid patients' in Lahore and to check the occurrence of *S. Typhi* in the drinking water of respective areas. This presence of *S. Typhi* will indicate whether water is the source of infection or not.

Received on 11-10-2021

Accepted on 22-04-2022

METHODOLOGY

Sample collection: After IRB permission, forty blood culture proven *S. Typhi* cases were questioned regarding variables such as dietary habits, and pets and detailed information about the source of water intake and whether the water was treated or not before consumption⁵. Then, a visit to the household and neighboring area of the cases was done and water samples were collected from tap⁴. From these 40 blood culture proven cases 120 water samples were taken. 82 water samples were taken randomly from different parts of Lahore. 500mL of water sample was collected in two 250mL sterile glass bottles. The glass bottles were transported to the microbiology laboratory in an icebox by maintaining a temperature of 4°C. The sample was analyzed as soon as it reached the laboratory⁵.

Sample processing: The 500mL water sample was divided into ten 50mL falcon tubes. Centrifugation at 3940rpm for 30 minutes was carried out of each falcon tube. The supernatant was removed and the sediment was added in the enrichment media (Selenite F Broth). The bottle was incubated for 8 – 10 hours at 37°C⁶. The sample was sub-cultured on the Xylose Lysine Deoxycholate medium and incubated at 37°C overnight. The isolated colonies were obtained after overnight incubation. All colonies were Gram stained. The colonies consisting of Gram negative rods were tested biochemically.

Catalase test was performed on all of the Gram negative rods. Then oxidase test and tube biochemical reactions (Citrate test, Triple Sugar Iron Test, Urease Test, Motility test, Indole Test, Methyl Red Test and Voges Proskauer Test) were performed. For each biochemical test a known positive and a known negative control was performed. Isolates tested positive for *Salmonella* species on Tube biochemical reactions were again tested via API20E for confirmation.

After the biochemical confirmation of the *Salmonella* species, Serotyping was performed for the serovars confirmation of the *Salmonella* species.

RESULTS

Total 202 water samples were tested. Growth upon subculture were observed in 153 water samples, implying a percentage of 76% indicating contamination and that drinking water was not fit for drinking. The water sample from the *Farooq Ganj*h area was tested

positive for *Salmonella* species, upon serotyping the isolates were confirmed as *Salmonella Paratyphi A* and *Salmonella Rubislaw*. No *Salmonella Typhi* was isolated from any water sample. A number of organisms were isolated from the water samples (Table 1) (Figure1).

Serratia species and *Enterobacter* Species were the most isolated organisms, then being the *Escherichia Coli*, *Citrobacter* species, *Pseudomonas* Species, *Proteus* Species, *Acinetobacter* species, *Klebsiella* Species and *Providencia* Species.

Table 1: The table illustrates the percentage of water samples which were positive for microbial contamination.

Name of Bacteria	Frequency	%age
<i>Escherichia coli</i>	24	15.59
<i>Salmonella Rubislaw</i>	1	0.54
<i>Salmonella Paratyphi A</i>	1	0.54
<i>Klebsiella spp.</i>	6	3.76
<i>Enterobacter spp.</i>	33	21.51
<i>Proteus spp.</i>	10	6.45
<i>Acinetobacter spp.</i>	8	5.38
<i>Providencia spp.</i>	1	0.54
<i>Serratia spp.</i>	34	22.04
<i>Citrobacter spp.</i>	18	11.83
<i>Pseudomonas spp.</i>	13	8.60
<i>Bacillus spp.</i>	5	3.23
Total	153	100

22% (41 isolates) and then the *Enterobacter* spp. 22% (40 isolates). The third most frequently isolated spp. was *E. coli* 16% (29 isolates). Others were *Citrobacter* spp. 12% (22 isolates), *Pseudomonas* spp. 9% (16 isolates), *Proteus* spp. 6% (12 isolates), *Acinetobacter* spp. 5% (10 isolates), *Klebsiella* spp. 4% (7 isolates), *Bacillus* spp. 3% (6 isolates) and *Providencia* spp., *Salmonella Paratyphi A* and *Salmonella Rubislaw* as 1%.

Figure 1: Pie chart representing the percentages of the various isolated organisms. This pie charts represent that the isolated *Serratia* spp

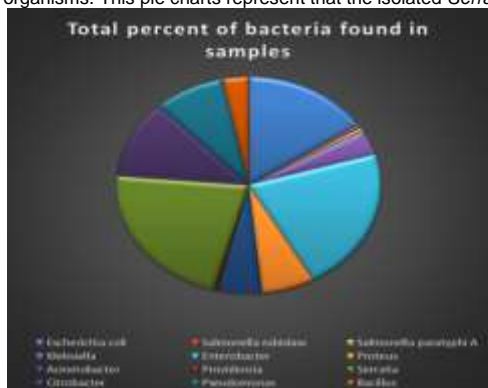


Figure 2: Geographical distribution of samples taken from the different areas of Lahore.



Indicates the location where the sample was taken from.

DISCUSSION

The water samples were collected from September 2020 to January 2021 from various areas of Lahore (Figure 2). The locality was surveyed and the hotspot areas were also targeted. The aim was to look for the presence *S. Typhi* in the drinking water supply of Lahore. Hence, enrichment media was included in the methodology before subculture upon a selective solid agar medium. This technique is a Gold Standard for the extraction of *Salmonella* species from fecal specimen. As the contamination of drinking water is majorly because of mixing of municipal sewerage / waste with the drinking water pipelines⁷.

The present study observed the presence of one *S. Paratyphi A* and one *S. Rubislaw* in the drinking water of Farooq Ganjh area, located in Lahore. The presence of *S. Paratyphi A* confirmed that the contaminated water was the main source of infection for the people residing in that area. The results were in accordance with study carried out in Tunisia, which stated that primarily the serovars *Paratyphi* and serovars other than *Typhi* are isolated from the environmental sources⁷. There are many studies conducted in the past which have isolated *Salmonella* species from drinking water. Hsu et al reported 10 *Salmonella* isolates from 116 drinking water samples¹. Potgieter observed the presence of *Salmonella* species from all of the household water samples from Limpopo province in all seasons⁸. In Nigeria, 4.35% were identified as *S. Typhi* from the drinking water of Ogbomoso North and 15.79% from Ogbomoso South by the similar technique⁹.

As discussed earlier, one *Salmonella Rubislaw* was also isolated from our samples. *S. Rubislaw* is a non-typhoidal *Salmonella* species and is the most common *Salmonella* serovars present in the water. *S. Rubislaw* is reported as the 11th most common *Salmonella* spp. causing disease¹⁰. This non-typhoidal *Salmonella* species is responsible for bacteremia in immunocompromised individuals and children. Other disease include self – limiting diarrhea¹¹. In Canada, a study concluded that 29 water samples out of 342 water samples were positive for *Salmonella* species. Within these positive samples *S. Rubislaw* was isolated from 21 water samples¹². In Ghana, 398 water samples were tested for the presence of coliforms. This study revealed that out of 392 water samples, 26 water samples were positive for *Salmonella* contamination. Within these 4 were *S. Rubislaw*¹¹. The presence of *S. Rubislaw* in all of the above studies indicated that ingestion of this contaminated water will be a health hazard for the humans¹².

Santo et al, studied the survival rate of four different species of *Salmonella* in drinking water. He concluded that the species remain viable even after 45 days and all the species respond to the addition of nutrients and growth¹³. Liu stated that in presence of adequate pH and temperature *Salmonella* spp. can survive in environmental sources for 30 days. As for *S. Typhi*, it can easily survive in water till 3 weeks and can easily be resuscitated upon addition of nutrients¹⁴. These studies conclude that *Salmonella* spp. can easily survive in drinking water for lengthy time span and upon enrichment these can easily be isolated.

Increasing number of patients of XDR *S. Typhi* were observed in Lahore in 2018 - 2020. Testing of the drinking water was done to check if it's the source of the disease. On testing no *S. Typhi* was isolated but two pathogenic *Salmonella* serovars were isolated. The incubation period of *S. Typhi* is 10 – 14 days and the patient may develop symptoms after 6 – 30 days of bacterial ingestion³. As there is a big time gap between the outbreak and the time at which water samples were taken, *S. Typhi* might not have survived in the water. Thus, it was concluded that the drinking water of Lahore might act as a single time source but is not acting as a constant source of typhoid caused by *S. Typhi* infection. Probably the main source of spread is mainly foodborne. Infected humans / carriers can give shelter to the bacteria for a long duration of time ranging from months to years.

These carriers / asymptomatic individuals shed the bacteria in the environment, contaminating food/water acting as a source of transmission of disease⁷. The carriers / food handlers working at the restaurants or food outlets may be the main source for the spread of the disease. For this purpose, further investigations should be done on this aspect.

Along with the isolation of the above mentioned *Salmonella* species, many of different species of bacteria were also isolated. *Serratia* species and *Enterobacter* Species were the most isolated organisms. *Escherichia Coli*, *Citrobacter* species, *Pseudomonas* Species, *Proteus* Species, *Acinetobacter* species, *Klebsiella* Species and *Providencia* Species were also isolated.

Serratia Species are commonly present in water and soil. *Serratia* has a medical importance as an opportunistic pathogen. It is associated with nosocomial infections, such as transfusion associated bacteremia and urinary tract infections. Endocarditis in drug addicts has also been reported¹⁵. In Germany, it was reported that out of 31 water samples 21% were positive for *Serratia* spp.¹⁶. In the present study 22% of water samples were positive for *Serratia* spp.

Enterobacter is also a common resident of water, soil and in the environment. *Enterobacter* is mostly causes urinary tract infections and nosocomial infections in the immunosuppressed individuals. In the recent years, bacteremia caused by *Enterobacter* has also been reported. Out of 4.7% bloodstream infections, 3.1% were caused by *Enterobacter*⁷. A very high percent of *Enterobacter* were also isolated from the water supplies of Mozambique. 36.36% of *Enterobacter cloacae* and 27.27% of *Enterobacter asburiae* were isolated¹⁷. 21.5% of water samples were positive for *Enterobacter* spp. in the present study indicating a health hazard.

Escherichia Coli is the most integral niche in the human intestinal flora. This is also termed as an indicator of fecal contamination in water. The *E. coli* is responsible for various intestinal and extra-intestinal infections in humans⁷. Different strains of *E. coli* are responsible for different features upon ingestion. The most common disease of *E. coli* is traveler's diarrhea. It is produced by the *Enterotoxigenic E. coli*. In South Africa the presence of *Enteropathogenic E. coli* and *Enteroggregative E. coli* in their drinking water supply were observed. These can result in the outbreak of diarrheal disease or critical illness in the immunocompromised individuals⁸. In the present study 15.6% of the water samples were positive for *E. coli*. The strain of *E. coli* was not identified as it was not the objective of our study but the presence of *E.coli* in our drinking water samples indicated fecal contamination of water.

Citrobacter, a Gram negative rod and a resident of human intestines, is also wide spread in environment especially water and soil. This dissemination in environment is mainly through sewerage water. It is also an intestinal pathogen due to its ability to produce enterotoxin⁷. *Citrobacter* has also been isolated from multiple brain abscesses but in neonates it is considered as an infrequent cause of bacterial meningitis¹⁷. 11.9% of *Citrobacter* spp. were isolated from the drinking water samples in the present study. Although *Citrobacter* is regarded as an opportunistic pathogen but its presence in the drinking water is worrisome.

Klebsiella is also a member of intestinal flora but it is also a pathogen for humans. It causes nosocomial infection. Nosocomial infections are spread through the hands of healthcare workers. *Klebsiella* is also responsible for pneumonia, urinary tract infections and liver abscesses. It also causes bacteremia and neonatal sepsis⁷. *Klebsiella* has been reported in the drinking water by a number of studies. A study conducted in Kathmandu, Nepal reported 18 samples positive for *Klebsiella* out of 100 ground water samples¹⁸. Only 3.76% of the water samples in our study were positive *Klebsiella* spp. posing a threat towards the wellbeing of the population.

The importance of the organisms such as *Citrobacter*, *Enterobacter*, *Klebsiella* and *Serratia* species in drinking water is that they also act as a source of antibiotic resistance spread¹⁹.

Proteus, a Gram negative rod, a member of *Enterobacteriaceae* is an inhabitant of soil and water. The existence of *Proteus* spp. in water is an important indicator of the fecal contamination. The main concern of *Proteus* presence in water is its highly antibiotic resistant strains. Although, *Proteus* is an opportunistic pathogen, it is responsible diarrhea, urinary tract infections and even wound infections²⁰. In the present study 6.45% of the water samples were positive for *Proteus* spp.

A study carried out in Nigeria, analyzed water samples from taps, storage tanks and distributional pipelines. Only one isolate of *Providencia* spp. was isolated from the total water samples²¹. Similarly, in our study only one sample was positive for *Providencia* spp.

Pseudomonas is a Gram negative rod, but doesn't belong from the family *Enterobacteriaceae*. These species are commonly found in the water sample as a contaminant. It has the ability to form biofilms within water pipelines. This causes the persistence of *Pseudomonas* species in water sample²². *Pseudomonas* spp. have been reported to impart taste and odor to the drinking water which might make it difficult for ingestion. In the present study, 8.6% of water samples were positive for *Pseudomonas* spp.

Non-coliforms such as *Acinetobacter*, was also isolated from the various areas of Lahore. *Acinetobacter* is mostly found in tap water and is also responsible for hospital acquired infections. The inhabitation of this organism in the water supplies is mostly linked with these infections. This was confirmed by a study carried in Tokai University Hospital²³. In the present study, 5.38% of the water samples were positive for *Acinetobacter*.

Bacillus species were the only Gram positive rods which were isolated. *Bacillus* spp. is the most common bacteria present in the environment. It occupies various environmental surfaces, from landscapes to soil and from soil to water²⁴. *Bacillus subtilis* is considered to act as a probiotic in humans. It is even thought that it should be added in water and beverages to increase the human health and decrease the age related illnesses²⁵. 3.23% of water samples were positive for *Bacillus* spp. in the present study.

CONCLUSION

It is concluded from the present study that drinking water of Lahore contained coliforms and non-coliforms which were widely distributed in the water supply of Lahore. This depicted faecal contamination of the drinking water. The presence of these pathogenic bacteria depicted that these might act as a source of infection for the whole population. Although, *S. Typhi* was not isolated but one sample was positive for *S. Paratyphi A* which represented that this serovars was responsible for Enteric fever in that area till samples were taken.

Acknowledgements: This work was supported by Department of Microbiology, University of Health Sciences, Lahore, Pakistan.

Conflict of interest: There is no conflict of interest among the authors.

Authorship contribution:

Fatima Tuz Zahra: Sample collection, methodology, software, data curation, writing – original draft, formal analysis, investigation, writing - review & editing and project administration.

Sidrah Saleem: Conceptualization, supervision, resources, funding acquisition.

Muhammad Imran: Supervision, writing review, validation.

Nida Javed: Sample collection, methodology, writing review.

Ayesha Ghazal: Methodology, writing review, investigation.

REFERENCES

- Hsu BM, Huang KH, Huang SW, Tseng KC, Su MJ, Lin WC, Ji DD, Shih FC, Chen JL, Kao PM. Evaluation of different analysis and identification methods for Salmonella detection in surface drinking water sources. Science of the total environment. 2011 Sep 15;409(20):4435-41.
- Admassu D, Egata G, Teklemariam Z. Prevalence and antimicrobial susceptibility pattern of Salmonella enterica serovar Typhi and

- Salmonella enterica serovar Paratyphi among febrile patients at Karamara Hospital, Jigjiga, eastern Ethiopia. SAGE Open Medicine. 2019;7:205031211983785
3. Gunn JS, Marshall JM, Baker S, Dongol S, Charles RC, Ryan ET. Salmonella chronic carriage: epidemiology, diagnosis, and gallbladder persistence. Trends in microbiology. 2014 Nov 1;22(11):648-55.
 4. Yousafzai MT, Qamar FN, Shakoor S, Saleem K, Lohana H, Karim S, Hotwani A, Qureshi S, Masood N, Rauf M, Khanzada JA. Ceftriaxone-resistant Salmonella Typhi outbreak in Hyderabad City of Sindh, Pakistan: high time for the introduction of typhoid conjugate vaccine. Clinical Infectious Diseases. 2019 Feb 15;68(Supplement_1):S16-21.
 5. Kovačić A, Huljev Ž, Sušić E. Ground water as the source of an outbreak of Salmonella Enteritidis. Journal of epidemiology and global health. 2017 Sep 1;7(3):181-4.
 6. Matrajt G, Lillis L, Meschke JS. Review of methods suitable for environmental surveillance of Salmonella Typhi and Paratyphi. Clinical Infectious Diseases. 2020 Jul 29;71(Supplement_2):S79-83.
 7. Cabral JP. Water microbiology. Bacterial pathogens and water. International journal of environmental research and public health. 2010 Oct;7(10):3657-703
 8. Potgieter N, Karambwe S, Mudau LS, Barnard T, Traore A. Human enteric pathogens in eight rivers used as rural household drinking water sources in the northern region of South Africa. International journal of environmental research and public health. 2020 Jan;17(6):2079.
 9. Bamigboye C, Amao J, Ayodele T, Adebayo A, Ogunleke J, Abass T et al. An appraisal of the drinking water quality of groundwater sources in Ogbomoso, Oyo state, Nigeria. Groundwater for Sustainable Development. 2020;11:100453
 10. Maurer JJ, Martin G, Hernandez S, Cheng Y, Gerner-Smidt P, Hise KB, Tobin D'Angelo M, Cole D, Sanchez S, Madden M, Valeika S. Diversity and persistence of Salmonella enterica strains in rural landscapes in the Southeastern United States. PloS one. 2015 Jul 1;10(7):e0128937.
 11. Dekker DM, Krumkamp R, Sarpong N, Frickmann H, Boahen KG, Frimpong M, Asare R, Larbi R, Hagen RM, Poppert S, Rabsch W. Drinking water from dug wells in rural Ghana—Salmonella contamination, environmental factors, and genotypes. International journal of environmental research and public health. 2015 Apr;12(4):3535-46.
 12. Jokinen C, Edge TA, Ho S, Koning W, Laing C, Mauro W, Medeiros D, Miller J, Robertson W, Taboada E, Thomas JE. Molecular subtypes of Campylobacter spp., Salmonella enterica, and Escherichia coli O157:H7 isolated from faecal and surface water samples in the Oldman River watershed, Alberta, Canada. Water research. 2011 Jan 1;45(3):1247-57.
 13. Santo Domingo JW, Harmon S, Bennett J. Survival of Salmonella species in river water. Current microbiology. 2000 Jun;40(6):409-17.
 14. Liu H, Whitehouse CA, Li B. Presence and persistence of Salmonella in water: the impact on microbial quality of water and food safety. Frontiers in Public Health. 2018 May 30;6:159.
 15. Rusin PA, Rose JB, Gerba CP. Health significance of pigmented bacteria in drinking water. Water science and technology. 1997 Jan 1;35(11-12):21-7.
 16. Reitter C, Petzoldt H, Korth A, Schwab F, Stange C, Hamsch B, Tiehm A, Lagkouravdos I, Gescher J, Hügler M. Seasonal dynamics in the number and composition of coliform bacteria in drinking water reservoirs. Science of The Total Environment. 2021 Sep 15;787:147539.
 17. Manhique GA, Hessel CT, Plessis EM, Lopes SM, de Oliveira Elias S, Tondo EC, Kortzen L. Prevalence of Enterobacteriaceae on Ready to Eat Salads, Drinking Water and Surfaces in Food Markets of Maputo, Mozambique. Journal of Food and Nutrition Research. 2020;8(1):63-73.
 18. Ghartimagar S, Khatri P, Neupane S, Joshi DR, Joshi TP. Evaluation of Ground Water Quality of Kathmandu Valley and Antibiotic Susceptibility Test against Klebsiella pneumoniae. Tribhuvan University Journal of Microbiology. 2020 Dec 27;7:83-90.
 19. Tanner WD, VanDerslice JA, Goel RK, Leecaster MK, Fisher MA, Olstadt J, Gurley CM, Morris AG, Seely KA, Chapman L, Korando M. Multi-state study of Enterobacteriaceae harboring extended-spectrum beta-lactamase and carbapenemase genes in US drinking water. Scientific reports. 2019 Mar 8;9(1):1-8.
 20. Drzewiecka D. Significance and roles of Proteus spp. bacteria in natural environments. Microbial ecology. 2016 Nov;72(4):741-58.
 21. Enaigbe AA, Ekhaie FO, Idemudia IB, Akpoka AO. Physicochemical and microbiological analyses of bacterial isolates from drinking water distribution systems of some higher institutions in Edo State, Nigeria. Journal of Applied Sciences and Environmental Management. 2019 Jun 18;23(5):909-15.
 22. Bertelli C, Courtois S, Roskiewicz M, Piriou P, Aeby S, Robert S, Lorent JF, Greub G. Reduced chlorine in drinking water distribution systems impacts bacterial biodiversity in biofilms. Frontiers in microbiology. 2018 Oct 23;9:2520.
 23. Umezawa K, Asai S, Ohshima T, Iwashita H, Ohashi M, Sasaki M, Kaneko A, Inokuchi S, Miyachi H. Outbreak of drug-resistant Acinetobacter baumannii ST219 caused by oral care using tap water from contaminated hand hygiene sinks as a reservoir. American journal of infection control. 2015 Nov 1;43(11):1249-51.
 24. Naderi Samani M, Jafaryan H, Gholipour H, Harsij M, Farhangi M. Effect of different concentration of profitable Bacillus on bioremediation of common carp (Cyprinus carpio) pond discharge. Iranian Journal of Aquatic Animal Health. 2016 Nov 10;2(2):44-54.
 25. Ayala FR, Bauman C, Cogliati S, Leñini C, Bartolini M, Grau R. Microbial flora, probiotics, Bacillus subtilis and the search for a long and healthy human longevity. Microbial Cell. 2017 Apr 3;4(4):133