

Modulation of Activity of Salivary Adrenomedullin by pH

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ABSTRACT

Background: Adrenomedullin is a multi-factorial peptide secreted by a range of human cells and is found in high levels in saliva. One potential role of Adrenomedullin is the maintenance of oral health through its antimicrobial activity.

Aim: To characterize how varying pH alters the antimicrobial effect of Adrenomedullin in short-term killing assays.

Study design: Experimental, Laboratory-based study.

Setting: Centre for Clinical and Diagnostic Sciences, Barts and The London, Queen Mary's School of Medicine and Dentistry, Queen Mary, University of London.

Method: Cultures of *Escherichia coli* and *Staphylococcus aureus* were used to demonstrate antimicrobial activity of Adrenomedullin at pH levels of 7.5, 6.5 and 5.5 by employing short-term bacterial killing assays.

Result: The results showed differences in the antimicrobial activity of salivary Adrenomedullin at different pH levels against the two different bacterial species used but were not significant. **Conclusion:** The results concluded that while Adrenomedullin has an antimicrobial effect, a greater range of pH and inclusion of specific oral microbes in the study will be helpful in investigating the hypothesis. However, it is likely that changes in pH in the oral cavity are important for the antimicrobial activity of salivary Adrenomedullin.

Keywords: Adrenomedullin (ADM), Minimum Inhibitory Concentration (MIC), Phosphate Buffered Saline (PBS)

INTRODUCTION

Adrenomedullin (ADM) is a 52 amino acid, multifunctional peptide produced by a wide variety of tissues and cells¹. It was originally isolated from extracts of pheochromocytoma using elevated platelet cAMP activity as an indicator². Previous studies have demonstrated the antimicrobial effect of ADM against a variety of normal skin, oral, gut and respiratory tract microflora^{3,4,5,6}. The Minimum Inhibitory Concentration (MIC) of whole ADM against *E. coli* and the Oxford strain of *S. aureus* has been found to be 12.5µg/ml (2.07µmol l⁻¹) whereas ADM fragments have exhibited different ranges of MIC individually. The mechanism of antibacterial action of ADM is believed to be by cell lysis via channel formation in the bacterial cell membranes⁷. Kapas and colleagues have detected the presence of ADM in whole saliva and various ductal salivary secretions of normal adults as well as from supernatants of salivary ductal cell lines⁴. A study carried out by Gröschl and colleagues suggests an important role of salivary ADM in the maintenance of oral health where it was shown to be involved in oral cell proliferation and antibacterial defense⁸. Gene expression profiling followed by real-time polymerase chain reaction analysis has detected ADM up-regulation in carious pulpal tissue. It has also been demonstrated that the gingival crevicular fluid ADM levels are higher in chronic periodontitis than periodontally health tissues⁹. *Porphyromonas gingivalis*, the major etiologic agent of chronic periodontitis, is killed by concentrations of Adrenomedullin above 500pmol/L¹⁰. ADM expression is elevated in response to polymicrobial attack and micro-environmental levels of ADM reach an effective antimicrobial peptide concentration¹¹. Ultrastructural analyses have shown cell wall disruption of microbes within 30 minutes of treatment with ADM in *E. coli* and abnormal septum formation with no cell wall disruption was observed in *S. aureus*⁵.

Dental caries is a microbial disease in which bacterial action on dietary fermentable carbohydrates produces acids which diffuse into the tooth causing demineralization of hydroxyapatite mineral. These acids diffuse through the plaque into the porous subsurface enamel dissociating to produce hydrogen ions thereby rapidly

lowering the pH in the mouth¹². The capacity of bacteria to produce acid in combination with growth at low pH is regarded as a virulence factor¹³. The onset and the rate of progression of dental caries are largely dependent on the action of dietary carbohydrates and microbes¹⁴. Additionally, it is also modulated by other factors such as the protective properties of saliva determined by the secretion rate, buffer capacity and antibacterial components¹⁵. The salivary pH normally varies from 6.0 to 7.5, with the most alkaline values obtained under stimulated flow rates¹⁶. In dental literature, the pH value that corresponds to the level of saturation is denoted as a fixed value of pH 5.5 and used as a threshold value for determination of demineralization of teeth in the oral cavity. The dependency of this value on the earlier mentioned determinants means that the critical pH in saliva is not constant, but a more dynamic variable varying around a mean pH of 5.5.

The aim of this study was to observe the antibacterial efficacy of human (whole) ADM against particular strains of *E. coli* and *S. aureus* at a range of pH given the dynamic nature of the pH of the mouth.

MATERIAL AND METHODS

E. coli 'NCTC 9001' and *S. aureus* 'oxford strain' were first incubated overnight in culture medium, followed by four hour (short-term) killing assays under a range of pH values of 5.5, 6.5 and 7.5 in triplicates and subsequently grown on agar plates under favorable conditions for colony counting. This study included two basic experiments which were repeated 3 times each using two different bacterial species.

Bacterial killing assays

Bacterial Growth Assays

Test Stock: A stock of 4 x Phosphate Buffered Saline (PBS) solution was made while maintaining ratio of PBS to distilled water at 1:3 and autoclaved. Different stocks of PBS solutions were made at pH 7.5, 6.5, 5.5 using a pH meter.

Culture Medium: Culture medium used was Tryptone Soy Broth (TSB). A stock of TSB was prepared by mixing 30g of TSB powder in 1L of distilled water which was then autoclaved and stored at room temperature.

Culture plates: Agar was prepared by mixing 40g of powder in 1L of purified water, autoclaved and subsequently immersed into a

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water bath at 55°C to prevent setting. Culture plates were poured in a laminar flow hood and were stored in a cold room at 4°C upon setting.

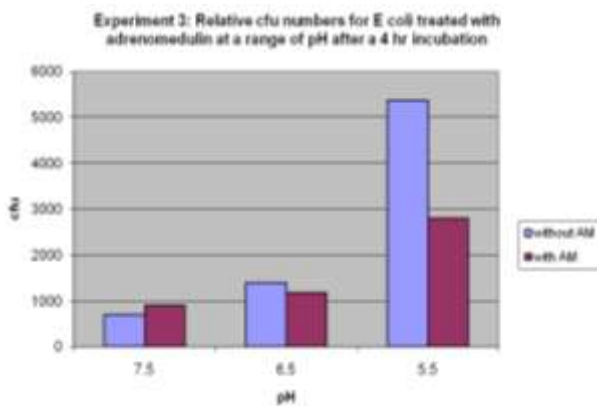
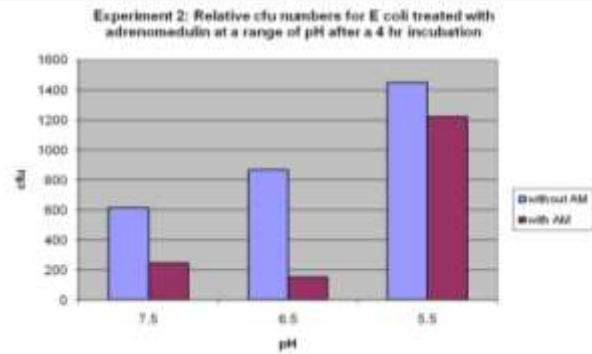
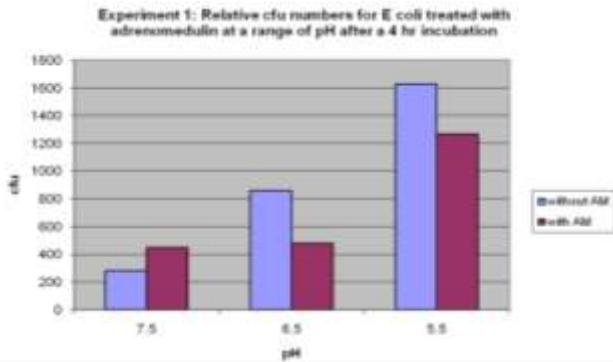
Bacterial Inoculum: 10ml of TSB was inoculated with bacterial strains employing the standard the standard protocol using wire loop and placed in an incubator shaker to allow overnight growth of bacteria at 37°C.

Serial Dilution and Control: Serial dilutions were done for PBS at pH 7.5, 6.5 and 5.5 with and without ADM up to 10⁻³ and also for control which was 10µl of PBS at pH 7.5 with addition of 10µl of inoculum. The control was not subjected to the 4 hour killing assay in incubator shaker

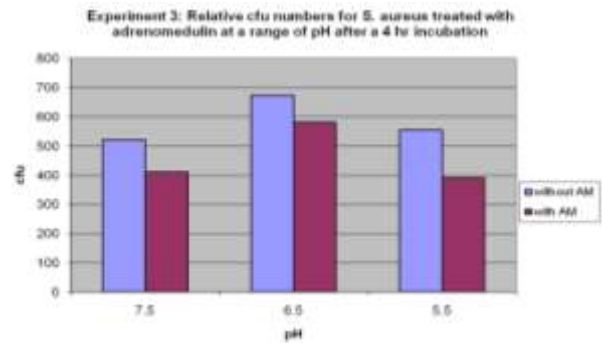
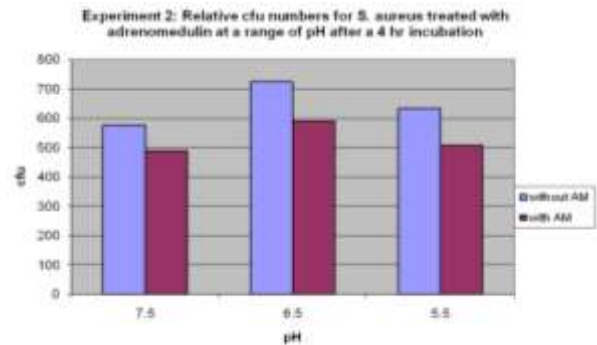
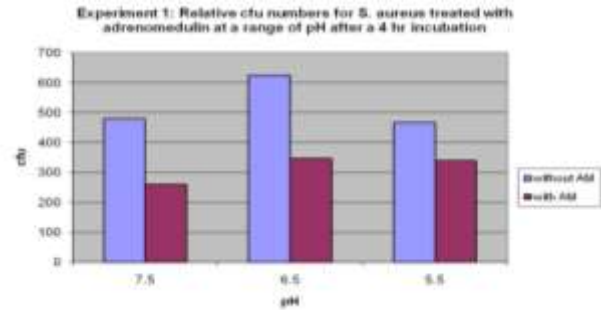
RESULTS

Killing Assays

E.coli: The experiments carried out in triplicates consistently showed decline in CFU in the presence of ADM indicating its antimicrobial action. There was a consistent downward trend of CFU at increased pH suggesting that this micro-organism favors lower pH for growth.

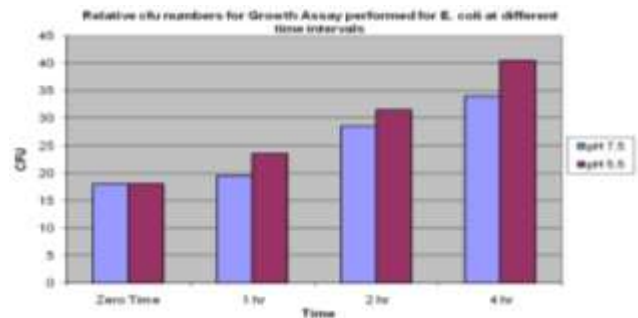


S. aureus: The results of all 3 experiments indicated a decline in CFU in the presence of ADM at all 3 pH levels signifying antimicrobial activity.

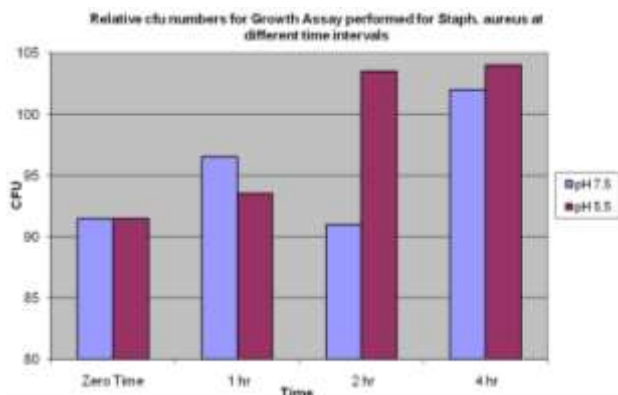


Growth Assays: These were carried out to gain an insight into the growth pattern of both bacterial species in relation to time and pH; and to compare the killing and growth assays for validation of findings obtained for the killing assays.

E. coli: Growth assays showed higher bacterial growth at pH 5.5 at all the time intervals thereby validating the findings of the killing assays.



S. aureus: Higher CFU numbers were observed with each time interval signifying bacterial growth. CFU numbers for the lower pH were more in comparison to the higher pH. CFU numbers were higher for *S. aureus* as compared to *E. coli* at the same Optical Density (OD).



Statistical Analysis: The results from different experiments would have been normalized according to the initial inoculum. Since this was not measured, the results therefore could not be expressed as the same way as the growth assays. Given that there seemed to be a pattern, we used a non-parametric test, (where the data does not fit a standard normal distribution) the Kruskal Wallis Test, to look for any significant association. No such association could be detected.

DISCUSSION

Our study further strengthens the findings of Allaker & colleagues¹⁷ in 1999 and Groschl⁸ in 2009 that ADM shows antimicrobial effects against Gram- positive and Gram-negative bacteria.

Bacterial growth inhibition depends largely on the concentration of the antimicrobial agent as well as the CFU of the bacteria used in experiments¹⁸. Growth inhibition assays could not be carried out in this study due to high cost of peptides. Reports in the literature suggest that electrostatic attraction between negatively charged bacterial cells and the positively charged ADM^{19,20}.

An interesting and evident finding in our study was that the CFU for *S. aureus* was noticeably higher than *E. coli*. Moreover, antimicrobial activity of ADM was not as efficient against *S. aureus* as against *E. coli*. It has been demonstrated that ADM has high affinity with *E. coli* lipopolysaccharide (LPS)²¹ and therefore, it is expected that Gram-negative bacteria would be more susceptible to killing than Gram-positive bacteria¹⁷. A similar trend was observed in this study.

CONCLUSION

The results concluded that salivary ADM has a definitive antimicrobial effect against the test organisms and changes are seen with varying values of pH supporting our hypothesis. However, a greater range of pH and inclusion of other bacterial species such as *P. gingivalis* will prove to be helpful in investigating the hypothesis and to further unravel the modulation of salivary ADM by varying pH.

Conflict of Interest: Nil

Authors' Contribution: **BS:** Concept & Design of Study, Principal Author, **ER:** Manuscript writing, **NK:** Statistical analysis, **SH:** Revisiting Critically, **OA:** Quality insurer & Data Collection, **MBS:** Interpretation of results, Final Approval of version **RZ:** Referencing & Final Approval of version

REFERENCES

- Nishikimi T, Kuwahara K, Nakagawa Y, Kangawa K, Nakao K. Adrenomedullin. In *Endocrinology of the Heart in Health and Disease* 2017 (pp. 41-58).
- Nishikimi T, Nakagawa Y. Adrenomedullin as a biomarker of heart failure. *Heart failure clinics*. 2017 Oct 7.
- Martínez-Herrero S, Martínez A. Adrenomedullin: Not Just Another Gastrointestinal Peptide. *Biomolecules*. 2022 Feb;12(2):156.
- Vázquez R, Riveiro ME, Berenguer-Daize C, O'kane A, Gormley J, Touzelet O, Rezai K, Bekradda M, Ouafik LH. Targeting adrenomedullin in oncology: a feasible strategy with potential as much more than an alternative anti-angiogenic therapy. *Frontiers in Oncology*. 2021;2678.
- Travers S, Martinerie L, Xue QY, Perrot J, Viengchareun S, Caron KM, Blakeney ES, Boileau P, Lombes M, Pussard E. Adrenomedullin: new inhibitory regulator for cortisol synthesis and secretion. *Journal of Endocrinology*. 2021 Aug 1;1(aop).
- Ferrero H, Larrayoz IM, Gil-Bea FJ, Martínez A, Ramírez MJ. Adrenomedullin, a novel target for neurodegenerative diseases. *Molecular Neurobiology*. 2018 Dec;55(12):8799-814.
- Ahsan H. Biomolecules and biomarkers in oral cavity: Bioassays and immunopathology. *Journal of Immunoassay and Immunochemistry*. 2019 Jan 2;40(1):52-69.
- Grant M, Kilsgård O, Åkerman S, Klinge B, Demmer RT, Malmström J, Jönsson D. The human salivary antimicrobial peptide profile according to the oral microbiota in health, periodontitis and smoking. *Journal of innate immunity*. 2019;11(5):432-44.
- Hussain QA, McKay IJ, Gonzales- Marin C, Allaker RP. Detection of adrenomedullin and nitric oxide in different forms of periodontal disease. *Journal of periodontal research*. 2016 Feb;51(1):16-25. Zudaire, E., Portal-Núñez, S. & Cuttitta, F. 2006. The central role of adrenomedullin in host defense. *J Leukoc Biol*, 80, 237-44.
- Domisch H, Skora P, Hirschfeld J, Olk G, Hildebrandt L, Jepsen S. The guardians of the periodontium—sequential and differential expression of antimicrobial peptides during gingival inflammation. Results from in vivo and in vitro studies. *Journal of Clinical Periodontology*. 2019 Mar;46(3):276-85.
- Gupta S, Bhatia G, Sharma A, Saxena S. Host defense peptides: An insight into the antimicrobial world. *Journal of oral and maxillofacial pathology: JOMFP*. 2018 May;22(2):239.
- Mukouyama C, Koike Y, Hirohara T. Transitional changes in the prevalence of dental caries in children and preventive strategies: a review of nationwide annual surveys in Japan. *Oral Health Prev Dent*. 2018 Jan 1;16(2):107-11.
- Arcari T, Feger ML, Guerreiro DN, Wu J, O'Byrne CP. Comparative Review of the Responses of *Listeria monocytogenes* and *Escherichia coli* to Low pH Stress. *Genes*. 2020 Nov;11(11):1330.
- Pitts NB, Zero DT, Marsh PD, Ekstrand K, Weintraub JA, Ramos-Gomez F, Tagami J, Twetman S, Tsakos G, Ismail A. Dental caries. *Nature reviews Disease primers*. 2017 May 25;3:17030.
- Maheswari E, Kumar RP, Arumugham IM, Sakthi DS, Lakshmi T. Evaluation of salivary flow rate, pH, buffering capacity, total calcium, protein, and total antioxidant capacity level among caries-free and caries-active children: A systematic review. *Journal of Advanced Pharmacy Education & Research* | Apr-Jun. 2017;7(2).
- Johnston TP. Anatomy and physiology of the oral mucosa. In *Oral Mucosal Drug Delivery and Therapy* 2015 (pp. 1-15). Springer, Boston, MA.
- Sigaud R, Dussault N, Berenguer-Daize C, Vellutini C, Benyahia Z, Cayol M, Parat F, Mabrouk K, Vázquez R, Riveiro ME, Metellus P. Role of the Tyrosine Phosphatase SHP-2 in Mediating Adrenomedullin Proangiogenic Activity in Solid Tumors. *Frontiers in oncology*. 2021;11.
- Loo YY, Rukayadi Y, Nor-Khaizura MA, Kuan CH, Chieng BW, Nishibuchi M, Radu S. In vitro antimicrobial activity of green synthesized silver nanoparticles against selected gram-negative foodborne pathogens. *Frontiers in microbiology*. 2018 Jul 16;9:1555.
- Lei J, Sun L, Huang S, Zhu C, Li P, He J, Mackey V, Coy DH, He Q. The antimicrobial peptides and their potential clinical applications. *American journal of translational research*. 2019;11(7):3919.
- Milardi D, Gazit E, Radford SE, Xu Y, Gallardo RU, Caffisch A, Westermark GT, Westermark P, Rosa CL, Ramamoorthy A. Proteostasis of islet amyloid polypeptide: a molecular perspective of risk factors and protective strategies for type II diabetes. *Chemical Reviews*. 2021 Jan 11;121(3):1845-93.
- Tsuruda T, Kato J, Kuwasako K, Kitamura K. Adrenomedullin: continuing to explore cardioprotection. *Peptides*. 2019 Jan 1;111:47.