The Novel Role of β -Aescin in Attenuating Isoniazid (INH)- Induced Hepatotoxicity in Rats

FARHEEN MALIK¹, SALAHUDDIN SHAIKH², ARSALAN AHMED UQAILI³, NAVAID KAZI⁴, FARZANA RAHIM⁵, SEHAR GUL⁶

¹Assistant Professor, Department of Pharmacology, Isra University, Hyderabad

^{2,5}Assistant professor, department of Physiology, Isra University, Hyderabad

³Assistant Professor, Department of Physiology, LUMHS, Jamshoro

⁴Professor, department of Physiology, Isra University, Hyderabad

⁶Assiatant Professor, Department of Physiology, Indus Medical College Tando Muhammad Khan

Corresponding author: Dr Arsalan Ahmed Uqaili, Email: drarsalan311@gmail.com

ABSTRACT

Objective: To investigate the hepatoprotective effect and underlying mechanisms of β -Aescin in Isoniazid (INH)-induced liver damage.

Methodology: This experimental study was carried out at the Pharmacology department and Postgraduate Laboratory of Isra University Hyderabad with the collaboration of Sindh Agricultural University Tando Jam. Forty adult Albino Wister rats, weighing between 200 and 250 grams were taken under a well-ventilated & hygienic environment at 26°C of room temperature and light/dark cycle of 12 hours and divided into 5 groups, each group containing 8 rats. After experiment animals were sacrificed via midline incision, the heart was identified and blood was drawn by cardiac puncture into a plain vial. The blood was allowed to clot and serum separated and stored at 20°C till further use. The liver was identified, removed and rinsed with normal saline and weighed. Serum levels of alanine aminotransferase (ALT), aspartate amino- transferase (AST) and bilirubin total were assessed using a commercially available kit. All the data were collected via study proforma, and analysis was done using SPSS version 26.

Results: A total of 40 animals after dividing in 5 groups as per study protocol were studied; to observe the hepatoprotective effect of β -Aescin in Isoniazid (INH)-induced liver damage. The body weight of the animal was significantly decreased in Isoniazid (INH) control group, experimental group C and D, while it was cured by β -Aescin 3.6 mg/kg. The findings are showing that the Isoniazid (INH) drug also significantly decreased the body weight and β -Aescin 3.6 mg/kg showed the significant protective effects in the body weight. The weight of the liver was significantly increased in Isoniazid (INH) group and experimental group C and D in contrast to the normal control (p-0.001), while experiment group E (β -Aescin 3.6 mg/kg) showed almost equal liver weight compared to the control normal group (p-0.345). β -Aescin 3.6 mg/kg showed the significant protective effects of hepatotoxicity induced by Isoniazid (INH), as the average of ALT, AST, total bilirubin and direct bilirubin were significant decreased almost near to control normally in the β -Aescin 3.6 mg/kg treated group (p-0.001).

Conclusion: As per study conclusion the β -Aescin 3.6 mg/kg was observed to be a protective agent against Isoniazid (INH)-induced hepatic toxicity. It was observed as the antioxidative and anti-inflammatory and antifibrotic to cure the liver. Hence, it can be used as a promising hepatoprotective agent.

Key words: Hepatotoxicity, tuberculosis drug, β-Aescin, effectiveness

INTRODUCTION

The liver is the major glandular and important organ in the body, and it is responsible for drug processing. It aids in the detoxification and excretion of several xenobiotics and medicinal substances.¹ Injury or destruction of its functions has serious consequences for the affected person's health.1 Hepatotoxicity is becoming a primary cause of death throughout the world, and its incidence is expanding at an exponential rate and the hepatotoxicity can be caused by several factors.² Hepatotoxicity is defined as liver malfunction or damage caused by an overabundance of medicines or xenobiotics. Hepatotoxins and hepatotoxicants are synthetic chemicals that cause liver injury. Exogenous hepatotoxicants include overdoses of some drugs, industrial chemicals, plants, and dietary supplements, among other things.^{2,3} Drugs undergo oxidation, hydrolysis, hydration, reduction, conjugation, condensation, or isomerization in the liver. Although the liver metabolizes medications, hepatotoxicity can occur if these systems are disrupted. Although drug-induced liver damage due to various medicines somewhat differs, the

clinical manifestations of INH-induced hepatic injury.⁴ One of the most serious side effects associated with anti-TB medications are drug-induced hepatotoxicity (DIH). Hepatic injury is usually asymptomatic and could only be discovered by detecting hepatocyte injury markers like (AST) aspartate aminotransferase and alanine aminotransferase (ALT). A transitory increase in serum alanine aminotransferase (ALT) occurs in about 10%-20% of cases who consume INH. The majority of patients adjust to it and whose serum ALT concentrations return to normal without treatment, but a small percentage of individuals (less than 1%-3%) acquire serious hepatic injury and even liver failure.^{6,7} The hepatotoxicity of INH has been studied using several animal studies, including rats, Nevertheless, no animal model has been established to mimic the human patterns of INH-induced liver injury.^{8,9} Hepatic damage is a significant condition that has ramifications on many levels, including specific patient health, pharmaceutical regulatory choices, and therapeutic development plans. Due to lack of complete cure, multiple unpleasant effects, decreased safety, and other factors, currently available medication

treatments are insufficient to meet clinical demand.¹⁰ As a result, the usage of herbal medications, which have a higher safety and efficacy profile, is becoming more popular. β -Aescin is the main active ingredient isolated from the Aesculus hippocastanum L horse chestnut tree.¹⁰ Furthermore, β -Aescin's anti-inflammatory and antioxidant properties have been shown to protect against endotoxin-induced hepatic injury.¹⁰ This study has been done to assess the hepatoprotective action of β -Aescin and underlying processes in Isoniazid (INH)-induced hepatic damage.

MATERIAL AND METHODS

This experimental study was carried out at the Pharmacology department and Postgraduate Laboratory of Isra University Hyderabad. Animal protocol was followed at the Animal House Department of Husbandry and Veterinary Sciences, Sindh Agricultural University Tando Jam. Forty adult Albino Wister rats, weighing between 200 and 250 grams were taken and divided into 5 groups, each group containing 8 rats. The animals were kept in plastic cages and equipped with plastic drinkers with nozzles of stainless steel and feed containers of stainless steel. They were free to access a standard chow diet and water before and following the experiments. The animals were kept under a well-ventilated & hygienic environment at 26°C of room temperature and light/dark cycle of 12 hours.

Animals were divided into five groups (8 animals in each group) as the following:

Group A (Control) were given normal chow diet along with clean water ad libitum.

Group B (Isoniazid (INH) control) group.

Group C (Experimental, β-Aescin 0.9 mg/kg treated group). **Group D** ((Experimental β-Aescin 1.8 mg/kg treated group))

Group E ((Experimental β -Aescin 3.6 mg/kg treated group).

The weights of the control and experimental animals were measured after the experiment. All the animals were sacrificed via cervical dislocation. The blood was obtained using the cardiac puncture procedure for biochemical examination. On an automatic modular analyzer, liver function tests (AST, ALT, and total bilirubin) were performed using a diagnostic kit approach. The livers of rats were removed, and gross morphological characteristics (weight and size) were recorded after washing using normal saline. A Petri dish was balanced at zero on an electronic precision balance (sartorex) to determine the hepatic weight. The liver size was determined using a measuring scale, and the livers were subsequently fixed in 10% formalin for histological investigation. Tissues were processed through a series of ethyl alcohol grades before being cleared in xylene. The tissues were treated using the paraffin embedding procedure to create paraffin blocks. Using a microtome, 4 micrometer sections were cut for slides and stained with hematoxylin and eosin for observation at 100 and 400 magnifications under a light microscope. All the data were collected via study proforma, and analysis was done using SPSS version 26.

RESULTS

A total of 40 animals after dividing in 5 groups as per study protocol were studied; to observe the hepatoprotective

effect of β -Aescin in Isoniazid (INH)-induced liver damage. The body weight of the animals was significantly decreased in Isoniazid (INH) control group, experimental groups C and D, while body weight was increased in the normal control group and experimental group E.

Table. 1 Mean of difference between initial and final body weight in all study groups n=40

all study groups n=+0				
Study groups		Pre to post experimental difference in body weight (grams)		
	N	Mean	Std. Deviation	
Normal control	8	15.21	3.144	
Isoniazid (INH) control	8	-19.17	7.264	
β-Aescin 0.9 mg/kg treated	8	-8.40	2.003	
β-Aescin 1.8 mg/kg treated	8	-3.84	3.324	
β-Aescin 3.6 mg/kg treated	8	2.54	2.891	

Table.2 Liver weight comparison of control and INH groups with experimental groups n=40

Groups	weight of liver (grams)		p-value	F-value
A vs B	4.23 <u>+</u> 0.175	7.11 <u>+</u> 0.645	0.001	
A vs C	4.23 <u>+</u> 0.175	5.48 <u>+</u> 0.560	0.001	
A vs D	4.23 <u>+</u> 0.175	5.37 <u>+</u> 0.399	0.001	
A vs E	4.23 <u>+</u> 0.175	4.66 <u>+</u> 0.400	0.345	40.475
B vs C	7.11 <u>+</u> 0.645	5.48 <u>+</u> 0.560	0.001	
B vs D	7.11 <u>+</u> 0.645	5.37 <u>+</u> 0.399	0.001	
B vs E	7.11 <u>+</u> 0.645	4.66 <u>+</u> 0.400	0.001	

Table.3 Lipid profile comparison	of	control	and	INH	groups	with
experimental groups n=40						

· · ·	ital groups n=+0		1			
Groups	ALT(IU/L)		p- value	F-value		
A vs B	45.88 <u>+</u> 9.465	206.54 <u>+</u> 18.790	0.001			
A vs C	45.88+9.465	152.03+9.257	0.001			
A vs D	45.88 <u>+</u> 9.465	91.57 <u>+</u> 9.756	0.001			
A vs E	45.88 <u>+</u> 9.465	72.06 <u>+</u> 10.020	0.001	233.606		
B vs C	206.54 <u>+</u> 18.79	152.03 <u>+</u> 9.257	0.001			
B vs D	206.54 <u>+</u> 18.79	72.57 <u>+</u> 9.756	0.001			
B vs E	206.54 <u>+</u> 18.79	55.06 <u>+</u> 10.020	0.001			
AST(IU/L	_)					
A vs B	44.59 <u>+</u> 7.315	170.88 <u>+</u> 19.533	0.001			
A vs C	44.59 <u>+</u> 7.315	128.14 <u>+</u> 6.286	0.001			
A vs D	44.59 <u>+</u> 7.315	83.19 <u>+</u> 3.675	0.001			
A vs E	44.59 <u>+</u> 7.315	71.25 <u>+</u> 6.037	0.001	190.531		
B vs C	170.88 <u>+</u> 19.53	128.14 <u>+</u> 6.286	0.001			
B vs D	170.88 <u>+</u> 19.53	83.19 <u>+</u> 3.675	0.001			
B vs E	170.88 <u>+</u> 19.53	71.25 <u>+</u> 6.037	0.001			
Total bilir	ubin					
A vs B	0.22 <u>+</u> 0.282	1.35 <u>+</u> 0.299	0.001			
A vs C	0.22 <u>+</u> 0.282	0.95 <u>+</u> 0.044	0.001			
A vs D	0.22 <u>+</u> 0.282	0.64 <u>+</u> 0.181	0.020			
A vs E	0.22 <u>+</u> 0.282	0.40 <u>+</u> 0.365	0.610	23.991		
B vs C	1.35 <u>+</u> 0.299	0.95 <u>+</u> 0.044	0.034			
B vs D	1.35 <u>+</u> 0.299	0.64 <u>+</u> 0.181	0.001			
B vs E	1.35 <u>+</u> 0.299	0.40 <u>+</u> 0.365	0.001			
Direct bilirubin						
A vs B	0.18 <u>+</u> 0.115	1.07 <u>+</u> 0.641	0.001			
A vs C	0.18 <u>+</u> 0.115	0.66 <u>+</u> 0.193	0.060			
A vs D	0.18 <u>+</u> 0.115	0.42 <u>+</u> 0.335	0.619			
A vs E	0.18 <u>+</u> 0.115	0.27 <u>+</u> 0.165	0.979	0.8552		
B vs C	1.07 <u>+</u> 0.641	0.66 <u>+</u> 0.193	0.145			
B vs D	1.07 <u>+</u> 0.641	0.42 <u>+</u> 0.335	0.005			
B vs E	1.07 <u>+</u> 0.641	0.27 <u>+</u> 0.165	0.001			

The findings are showing that the Isoniazid (INH) drug also significantly decreased the body weight and β -Aescin 3.6 mg/kg showed the significant protective effects in the body weight. Table.1 Weight of the liver was significantly increased in Isoniazid (INH) group and experimental group C and D in contrast to the normal control (p-0.001), while experiment group E (β -Aescin 3.6 mg/kg) showed almost equal liver weight compared to the control normal group (p-0.345). Table.2 β -Aescin 3.6 mg/kg showed the significant protective effects of hepatotoxicity induced by Isoniazid (INH), as the average of ALT, AST, total bilirubin and direct bilirubin were significant decreased almost near to control normally in the β -Aescin 3.6 mg/kg treated group (p-0.001), the results shown in table.3

DISCUSSION

In this study, hepatotoxicity by acute administration of isoniazid was investigated; it could indeed be ameliorated by treatment with Aescin. With confidence and a rigorous search of the medical literature, this study being reported from the Pharmacology Department of Isra University. Isoniazid (INH) is a first-line drug of anti-tuberculosis drug regimens of the current era. INH is notorious of its liver damaging potential, culminating in severe acute liver failure and even death may occur in a few patients. Instead of this, it is being used in clinical practice because of no alternative effective agents against **Mvcobacterium** drug tuberculosis.¹¹⁻¹⁴ The β -Aescin exerts anti-inflammatory effects through its inhibitory effects on the histaminergic and serotonergic receptors.¹⁵ The β-Aescin inhibits the Phospholipase A2 (PLA2) enzyme that generates the arachidonic acid derived inflammatory mediators. The β-Aescin inhibits the inflammatory reaction in hypoxia induced human endothelial cell injury. This is the major anti-inflammatory pathway through which β-Aescin exerts clinical response and alleviation of edema and inflammation.¹⁶ Liu et al¹⁷ conducted study on the in vitro effects of β-Aescin against the periodontal ligament inflammation. They found a significant inhibitory effect on the expression of Toll-like receptor (TLR). The proinflammatory cytokines tumor necrosis factor-a (TNF-a), interleukin-1ß (IL-1ß) and IL-6 were reduced in a lipopolysaccharide induced inflammatory animal model.¹⁸ Aescin also improved the liver histology and protected the liver parenchyma against INH injury. The findings of our study are consistent with the previousthose of previous studies.^{19,20} In a study by Wei et al the hepatoprotective effects of CoQ10 in isoniazid fed rats has been observed as determined by the liver enzymes and histology, serum bilirubin, ALT, AST, LDH, ALP, GGT, Albumin, GPX, TNF-a . Similar effects were obtained by concomitant Aescin + INH in our study. Wang et al²¹ has reported an experimental rat study for the effectiveness of Aescin against the indomethacin induced gastric ulcer. The findings support the histological protective effects of aescin similar to this study. However, they used indomethacin instead of Isoniazid that is not similar to this study. Ezberrci F et al²² reported the effects of Aescin in the pile/hemorrhoids. They reported that aescin is effective in vanishing the piles. The findings of the above study is a worth regarding the pharmacological effect of aescin as also observed in the present study. Gupta SC et al²³

reported the down regulation of tumor necrosis factor and other proinflammatory biomarkers by polyphenols of Aescin. The finding is in agreement with the present study as we have observed similar finding of down regulation of TNF-α. However; other pro- inflmmatory mediators were not analyzed in the present study that is in disagreement. Krause et al²⁴ and Roman et al²⁵ reported anti-fungal activity of Aescin on dermatophytes in vitro. This finding supports the present study that the Aescin has pharmacological effects. Zhang et al²⁶ studied the effects of aescin on the Tumor Necrosis Factor in Human Immunity. Above the previous study reported the Aescin down regulates the effects of TNF-a. This pharmacological finding was in agreement with the present study. However, other cytokine markers were also studied by Zhang et al and those were not analyzed in the present study; hence, other findings of cytokines are inconsistent. Elmas et al²⁷ analyzed the effects of Aescin on Diabetic nephropathy in a streptozotocin-induced diabetic rat model. They reported the antioxidant (GPX) was increased and TNF- α was decreased. They also reported the histoprotective effects of Aescin on rat kidneys. The present study used an experimental model of liver histology that was ameliorated by Aescin. The histological effects of Elmas et al²⁷ on the kidney, although contradict to the liver histology, but still support the present study.

CONCLUSION

After the study β -Aescin β -Aescin 3.6 mg/kg was observed to be protective agent against Isoniazid (INH)-induced hepatic toxicity. It was observed as the antioxidative and anti-inflammatory and antifibrotic to cure the liver. Hence, it can be used as a promising hepatoprotective agent. Hepato-protective role of Aescin might have been mediated partly due to its anti-inflammatory effect and anti-oxidative mechanisms as has been observed by Glutathione peroxidase and Tumor necrosis factor- α .

REFERENCES

- Singh H, Sidhu S, Chopra K, Khan MU. The novel role of βaescin in attenuating CCl4-induced hepatotoxicity in rats. Pharmaceutical biology. 2017 Jan 1;55(1):749-57.
- 2. Alshehri MM, Amjad MW, Mudawi MM. Drugs-Inducing Hepatotoxicity. AJPRHC. 2020 Sep 1;12(3):148-56.
- Singh A, Bhat TK, Sharma OP. Clinical Biochemistry of Hepatotoxicity. J Clinic Toxicol S4: 001. doi: 10.4172/2161-0495. S4-001 J Clinic Toxicol Clinical Pharmacology: Research & Trials ISSN: 2161-0495 JCT, an open access journal. vitro Systems. 2011.
- Metushi I, Uetrecht J, Phillips E. Mechanism of isoniazidinduced hepatotoxicity: then and now. BJCP 2016;81(6):1030-6.
- 5. Lei S, Gu R, Ma X. Clinical perspectives of isoniazid-induced liver injury. Liver Research 2021; 1;5(2):45-52.
- Centers for Disease Control and Prevention (CDC. Severe isoniazid-associated liver injuries among persons being treated for latent tuberculosis infection-United States, 2004-2008. MMWR. Morbidity and mortality weekly report. 2010 Mar 5;59(8):224-9.
- Wang P, Pradhan K, Zhong XB, Ma X. Isoniazid metabolism and hepatotoxicity. Acta pharmaceutica sinica B. 2016;ep 1;6(5):384-92.
- 9. U.A. Boelsterli, K.K. Lee. Mechanisms of isoniazid-induced idiosyncratic liver injury: emerging role of mitochondrial stress. J Gastroenterol Hepatol, 29 (2014), pp.

- Singh H, Sidhu S, Chopra K, Khan MU. The novel role of βaescin in attenuating CCl4-induced hepatotoxicity in rats. Pharmaceutical biology. 2017 Jan 1;55(1):749-57.
- Khan SR, Morgan AĞ, Michail K, Srivastava N, Whittal RM, Aljuhani N, et al. Metabolism of isoniazid by neutrophil myeloperoxidase leads to isoniazid-NAD+ adduct formation: a comparison of the reactivity of isoniazid with its known human metabolites. Biochemical pharmacology. 2016;106:46-55.
- 12. Metushi IG, Nakagawa T, Uetrecht J. Direct oxidation and covalent binding of isoniazid to rodent liver and human hepatic microsomes: humans are more like mice than rats. Chemical research in toxicology. 2012;25(11):2567-76.
- Singh H, Sidhu S, Chopra K, Khan M. The novel role of βaescin in attenuating CCl4-induced hepatotoxicity in rats. Pharmaceutical biology. 2017;55(1):749-57.
- Albus U. Guide for the Care and Use of Laboratory Animals (8th edn). SAGE Publications Sage UK: London, England; 2012.
- Matsuda H, Li Y, Murakami T, NINOMIYA K, YAMAHARA J, YOSHIKAWA M. Effects of escins Ia, Ib, IIa, and IIb from horse chestnut, the seeds of Aesculus hippocastanum L., on acute inflammation in animals. BPB. 1997;20(10):1092-5.
- Dudek-Makuch M, Studzińska-Sroka E. Horse chestnut– efficacy and safety in chronic venous insufficiency: an overview. Revista Brasileira de Farmacognosia. 2015;25(5):533-41.
- 17. Liu S, Wang H, Qiu C, Zhang J, Zhang T et al. Escin inhibits lipopolysaccharide-induced inflammation in human periodontal ligament cells. Molecular medicine reports. 2012;6(5):1150-4.
- Xin W, Zhang L, Fan H, Jiang N, Wang T, Fu F. Escin attenuates acute lung injury induced by endotoxin in mice. European Journal of Pharmaceutical Sciences. 2011;42(1-2):73-80.

- Domanski D, Zegrocka-Stendel O, Perzanowska A, Dutkiewicz M, Kowalewska M, Grabowska I, et al. Molecular mechanism for cellular response to β-escin and its therapeutic implications. PloS one. 2016;11(10):e0164365.
- 20. Güney G, Kutlu H, Işcan A. The Apoptotic Effects of Escin in The H-Ras Transformed 5RP7 Cell Line. PR. 2013;27(6):900-5.
- 21. Wang T, Zhao S, Wang Y, Yang Y, Yao L, Chu L, et al. Protective effects of escin against indomethacin-induced gastric ulcer in mice. Toxicology mechanisms and methods. 2014;24(8):560-6.
- 22. Ezberci F, Ünal E. Hemoroidal Hastalığın Tedavisinde Aesculus Hippocastanum (Aescin, At Kestanesi) Kullanımı: Derleme. 2018.
- 23. Gupta SC, Tyagi AK, Deshmukh-Taskar P, Hinojosa M, Prasad S, Aggarwal BB. Downregulation of tumor necrosis factor and other proinflammatory biomarkers by polyphenols. Archives of biochemistry and biophysics. 2014;559:91-9.
- Krause H, WEINERT V. Fungistatic effect of aescin on dermatophytes in vitro. Arzneimittel-Forschung. 1970;20(5):703-5.
- Franiczek R, Gleńsk M, Krzyżanowska B, Włodarczyk M. β-Aescin at subinhibitory concentration (sub-MIC) enhances susceptibility of Candida glabrata clinical isolates to nystatin. Medical mycology. 2015;53(8):845-51.
- Zhang L, Yao CH. The physiological role of tumor necrosis factor in human immunity and its potential implications in spinal manipulative therapy: a narrative literature review. Journal of chiropractic medicine. 2016;15(3):190-6.
- 27. Elmas O, Erbas O, Yigitturk G. The efficacy of Aesculus hippocastanum seeds on diabetic nephropathy in a streptozotocin-induced diabetic rat model. Biomedicine & Pharmacotherapy. 2016;83:392-6.