

Sensitivity of Plasma Micrnas as a Probable Non-Small Cell Lung Cancer Biomarker and Its Significance

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ABSTRACT

Non-small cell lung cancer (NSCLC) is the chief origin of death due to cancer. The development of a less invasive diagnostic technique for NSCLC, especially at the beginning, can improve outcomes. By means of microarray platforms, we formerly diagnosed 12 microRNAs (miRNAs) abnormally expressed in primary cancer cells associated with early NSCLC.

Objective: In this study, we will extend previous studies to determine if miRNAs may be beneficial as a NSCLC potential plasma biomarker.

Methods: We primarily confirmed miRNA expression from PCR of 32 stage one NSCLC patients with lung tumor and plasma specimens were assessed, then assessed the investigative value of plasma miRNAs in 61 patients of NSCLC and 30 normal subjects. It was confirmed that the alteration of MiRNA expression influences the regeneration of neoplastic tumors. MiRNAs were steady and reliable in plasma measurements. The cohort consisted of 21 men and 11 women. Their age ranges from 47 to 80 years. AC tumors were classified in 17 and SCC in 15 patients. Smoking packets use in these patients were 39 ± 28 . In addition, IV blood were taken from healthy subjects with matched data distribution to studied group as age, race, sex and smoking, and aided as a control to evaluate fluctuations in plasma in cancer patients.

Results: Five out of 12 miRNAs showed a significant change of level in plasma and the corresponding tumor tissue (all $r > 0.850$, all $P < 0.05$). Four genes (miRNA-126, 21, 486-5 and 210p) set the best logistic regression pattern, with 87% sensitivity and 97% of specificity of at the time of NSCLC in differentiating from healthy patients. Moreover, the miRNA genes generated a sensitivity of 79.41% and 92.59% of specificity in patients diagnosed with SCC and AC. Moreover, genes have specificity of 96.67% in the analysis of lung adenocarcinoma and squamous cell carcinoma. (P less than 0.05). A change in square miRNA expression may offer potential NSCLC blood-derived biomarker.

Conclusion: Finally, we show that miRNA expression identified from surgical tumor tissue can be easily and correctly determined in plasma. Though, prominently, a plasma miRNA recognition panel would be cast-off as a less invasive analytic method for NSCLC, including long-term adenocarcinoma. However, the presence of an independent potential biomarker requires further verification.

Keywords: Lung cancer; Diagnosis; Plasma; Microna; Qrt-PCR

INTRODUCTION

Non-small cell lung cancer (NSCLC) is the utmost communal lung cancer type and is the chief cause of mortality in the USA and around the world. NSCLC chiefly comprises of 2 main forms of histology: squamous cell carcinoma (SCC) and adenocarcinoma (AC). NSCLC is often detected in last stages, ensuing 14% 5-year survival rate, approximately. In contrast, the 5-year endurance percentage of stage I NSCLC positive individuals receiving successful treatment may be up to maximum of 83%. So, early detection of NSCLC can decrease death ratio¹⁻³. Prompt recognition of NSCLC on X-ray and sputum cytology has less sensitivity. Bronchoscopy is great for detecting central occurring tumors of the lungs. Though, it's an invasive procedure. Though computed tomography offers good anatomical information and can perceive noninvasively small-size NSCLC, better sensitivity is associated with overdiagnosis⁴. Therefore, the development of minimally invasive techniques using the latest advances in molecular genetics is clinically important for the early diagnosis of NSCLC⁵. Blood plasma is obviously the best option for developing these diagnostic

markers. Many molecular alterations of specific tumors are recognized in serum or plasma and have been revealed to be a biomarker in lung cancer patients⁶⁻⁷. However, none of the markers tested so far has achieved the features allowing for the NSCLC diagnosis. MicroRNAs (miRNAs) have revealed now discoveries in the molecular analysis of malignancy⁸. Proven studies reports that the blood circulating miRNAs, are helpful for detection of tumors including lung cancer, can be used as cancer biomarkers. E.g., recently, miRNA serum signatures have been recognized that can forecast the endurance of patients affected with NSCLC. While it is postulated that measuring miRNA expression in plasma / serum is a promising approach for the diagnosis of lungs tumor, the idea entails more research to validate its probable clinical benefit⁹⁻¹⁰. By means of microarray platforms, we newly recognized twelve miRNAs whom if abnormally expressed in human tumor tissues exhibited initial NSCLC staging¹¹. So, now we extend this research based on previous studies by examining whether miRNAs can be found in plasma and cast-off as less invasive biomarkers of NSCLC. First, we established the miRNA findings in pairs of surgical tissues

and specimens of plasma from independent cases of NSCLC by means of quantitative Real-time qRT-PCR. We then analyzed plasma miRNA expression in a cohort of 61 patients of NSCLC and 32 healthy individuals to evaluate the analyticalworth of NSCLC. Studies have shown that changes in plasma miRNA expression have the potential to provide blood-derived biomarkers for NSCLC¹².

MATERIAL AND METHODS

There are two phases of this study. In the 1st phase, we were all able to confirm whether the miRNAs changes are related with NSCLC stage I, the abnormal appearance of which could be detected in surgical independent tissues and whether the miRNAs change in the plasma matched appropriately with the equivalent tumors. So, we gained the paired tissues of lung tumor, plasma specimens and noncancerous lung tissues of 32stage I patients of NSCLC (Table 1).

Table 1 Shows 32 Stage-I NSCLC Cases Matched with 29 Healthy Subjects and Their Clinical and Characteristic Features

	32 cases of NSCLC	29 Healthy smokers
Age, median (s.d.)	67 (7.1)	68.2 (6.4)
Gender		
Male	11 (34.4%)	20 (69%)
Female	21 (65.6%)	9 (31%)
Status of Smoking		
Pack per years, median (s.d.)	36.4 (24.2)	36.9 (26.8)
Stage		
Stage I	32 (100%)	
The Histological types		
Adenocarcinoma	17 (53.1%)	
Squamous cell carcinoma	15 (46.9%)	

The study consisted of 21 men and 11 women. Their age ranges from 47 to 80 years. AC tumors were classified in 17 and SCC in 15 patients. The smoking packets use in these patients were 39 ± 28 per year. In addition, IV blood were taken from 29 healthy subjects with matched data distribution to studied group as age, race, sex and smoking, and aided as a control to evaluate fluctuations in plasma in cancer patients. The test was performed according to the study protocols. Blood samples from cancer patients were taken prior to anesthesia induction, prior to surgical procedure. No one was given adjuvant radiotherapy or chemotherapy preoperatively.

The 2nd phase discusses the diagnostic worth of miRNAs in NSCLC plasma. Blood samples were obtained according to the international protocols. NSCLC samples were collected from 61patients of SCC and AC and 30 healthy people (Table 2) by NSCLC at various stages and histological types. Of the cancer patients,22 were female, and 39 were male.

A total of 14 have NSCLC stage I cancer, stage II in 17 patients, stage III in 14 subjects, and stage IV in 16 cases. The diagnosis was madehistologically after thoracotomy and bronchoscopic biopsy according to the WHO strategy. The classification of NSCLC phases is grounded on the WHO classification and International Staging of Lung Cancer System. Differences in race, age, smoking and gender between healthy control sand NSCLC patients was not significant. Peripheral blood (10 ml) was withdrawn EDTA coated tubes. The cancer patients' blood was taken at the time of the primary consultation, prior to the final surgical treatment and / or adjuvant treatment. The samples were collected and processed at 1300 xg for two hours by centrifugation at four degree for 10 minutes. Plasma was relocated to a fresh tube and until use were preservedat 80 ° C.

RNA was taken out from the tissue samples by means of the mirVana miRNA isolation kit. 1.33 ulof the solution of cDNA was augmented in twenty microliterper ml of the mixtures. PCR quantitative was accomplished on a Bio-Red IQ5 RT multi-color spot detection system. All assays were accomplished in triplicate, and two inter-plate controls and a no template control were achieved in each trial. The expression levels in MiRNA were considered using the comparative cycle threshold (CT) method. The fold variation was considered using theequation of 2^{ΔΔCt}.

Table II. Shows 61 NSCLC Cases Matched with 30 Healthy Subjects and Their Clinical and Characteristic Features

	61 NSCLC cases		30 Healthy subjects
	27 SCC patients	34 AC patients	
Age, median (s.d.)	68.6 (8.9)	69.2 (8.4)	67.2 (7.9)
Sex			
Male	17 (63%)	22 (64.7%)	19 (63.3%)
Female	10 (37%)	12 (35.3%)	11 (36.7%)
Status of Smoking			
Pack per years, median (s.d.)	35.8 (28.1)	31.8 (21.3)	31.6 (28.5)
Stage			
I	8 (29.6%)	6 (17.6%)	14
II	8 (29.6%)	9 (26.5%)	17
III	7 (25.9%)	7 (20.6%)	14
IV	4 (14.8%)	12 (35.3%)	16

Sensitivity and Specificity of the qRT-PCR test for the MiRNAs quantification:

Plasma was taken from 12 healthy people and alienated into 4 portions of 350 ml. The 1st aliquots of each sample were isolated on a day first for RNA isolation, the rest were stored at 20 ° C and made at 3rd, 7th and 30th days. The 1st two aliquots of every sample were incubated with different concentrations of RNase A (Qiagen). (0.1 and one mg / ml) for twenty hours correspondingly. One-third of all specimen were taken as control and used for treatment. Additionally, it was introduced into plasma treated samples with the synthesized miR238 RNase A cell (Qiagen) at various absorptions and without exemplary treatment. Endogenous miRNA expression and spiked miRNA expression were assessed in parallel samples to qRT-PCR, respectively. Furthermore, the plasma of the remaining healthy people was alienated into 3 portions. The 1st aliquots of every sample were repeated until freezing for 5 cycles, and the remaining 2 aliquots were kept in the refrigerator at 801C for three and seven days, correspondingly. The endogenous miRNAs expression in these samples was also calculated. To evaluate the accuracy of the qRT-PCR assay for quantifying miRNA in plasma, the CEL-Mir-238 in synthesized form was diluted in RNA isolated form, respectively, from 5 denaturated samples of plasma. The primers precise qRT-PCR to cel-miR-238 was achieved in the samples. Sensitive plasma samples with CEL-Mir-238 in synthesized form were tried in parallel and cast-off as controls. The variants of miR-486 which were synthesized dilute in DEPC water, followed by polyadenylation and reverse transcription. Independent qRT-PCR reactions were performed for each Mir-486 variant whether in each reaction there were specific PCR primers to solitary one of the two variations. To quantify plasma miRNAs to govern the dynamic range and sensitivity of qRT-PCR, RNA was 1st taken from the 5 healthy individuals' plasma and then dilute in DEPC water by 10 folds. Expression of twelve miRNAs was evaluated by qRT-PCR in consecutively samples in diluted form. All trials were tripled and repeated for 3 times.

Pearson's correlation analysis was used to evaluate the relation amid expression of tumor tissue in miRNA with plasma paired specimens. The examination was also used to evaluate the association amid plasma miRNA expression and the patients' demographic characteristics and healthy subjects. Clinicopathological analyses were taken as a reference standard to determine the specificity and sensitivity of gene therapy. The AUC analysis and receiver-operator characteristic curve was used with some precision to regulate each miRNA accuracy and to know each miRNA cut-off value in a particular sample and specificity percentage. The constrained parameters of Logistic regression models and comparable to minimum selection operator and absolute shrinkage were pragmatic to select an ideal potential biomarkers panel. All analyzes, counting the Wilcoxon's test, correlation coefficient, ANOVA, student text and logistic regression were achieved with log-transformed data. All values of the P are two sided and the less than 0.05 value of P was also statistically significant.

RESULTS

Previously, microarray techniques were applied to examine primary lungs tumor and the expression variations we identified with 12 miRNAs were related with NSCLC stage I. MiRNAs are miRNAs-126, 21, 139, 145, 200b, 182, 210, 205, 375, 486-5p, 708 and 429. To govern whether miRNA abnormalities could be determined in tumor independent tissues we evaluated qRT-PCR miRNA expression from 32 stage I NSCLC tissues and comparison was made with the non-cancerous tissue. All MiRNAs showed double or less tumor tissue expression compared to the corresponding non-carcinogenic substances (all after 0.001) (Table 3).

Table III shows 32 Stage-I NSCLC cases and their MiRNAs differentially expressed versus noncancerous paired specimens of lung tissue by qRT-PCR

miRNAs	Log2 (fold)* (mean±s.d.)	P-value
miR-126	2.86785679±0.24785434	0.00028423
miR-21	2.68376890±0.27980678	0.00072867
miR-139	2.14678230±0.20696445	0.00015965
miR-145	2.47896724±0.21985626	0.00052779
miR-205	2.87982563±0.26758932	0.00058936
miR-182	2.22678450±0.28975487	0.00027005
miR-210	2.31678590±0.31235786	0.00076259
miR-200b	2.16779289±0.24567893	0.00032789
miR-429	2.01045975±0.31143657	0.00023868
miR-375	2.85620858±0.26895849	0.00057949
miR-708	2.27832248±0.32256860	0.00055094
miR-486-5p	2.86749864±0.28656724	0.00043332

To discover an internal control that can measure reliable miRNAs plasma expression, we assessed C_t qRT-PCR values of RNU6B and miR16 in all NSCLCs in plasma of healthy individuals. Both genes have formerly remained anticipated as a potential control for the internal normalization of quantitative miRNAs. To standardize the variance in experimental performance between various samples, all qRT sequences are derived from a given volume and the equal RNA amount or a fixed RNA volume of initial plasma (350 ml) was taken. Additionally, we compare the C_t endogenous RNU6B and Mir-16 values with the spiked cel-miR-238. There was no significant difference in cancer patients in newly ready C_t plasma for CT RNU6B (P = 0.418) and Mir-16 (P = 0.464) values.

Table IV Shows Altered miRNA Expressions Prevalence in The Plasma Samples and Paired Tumor from 32 Stage-I NSCLC Cases

miRNAs	The number of individuals with transformed expressions of miRNA (%)	Positive cases in Plasma (%)
	Positive cases of Tumor, (%)	
miR-182	30/32, (93.75)	21/32, (65.63)
miR-126	32/32, (100)	22/32, (68.75)
miR-486-5p	32/32, (100)	24/32, (75.00)
miR-210	29/32, (90.63)	20/32, (62.50)
miR-21	31/32, (96.88)	23/32, (71.88)

NSCLC-associated miRNA expression was reliably measured in plasma:

To govern if there were twelve miRNAs in the plasma, we evaluated its expression in the plasma of 12 healthy persons done by qRT-PCR. The total

miRNAs tested had less than 32C_t values in every sample, representing that the miRNAs were in the plasma and can be demonstrable. We then promoted the endogenous miRNAs stability in archival samples of plasma. The plasma samples aliquots were stored for one, three, seven and thirty days at 20 ° C. The bioanalyst exhibited that the total RNA gradually decreased with the increase in 18S and 28S peaks from day 1st to 30th day.

Though, no effect on the miRNA expression level was observed, which was confirmed by qRT-PCR in the similar sample.

Since plasma had an increase level of RNase activity, we measured miRNA expression treated in the samples with or without RNase A. To evaluate the accuracy of the quantification qRT-PCR of the plasma miRNAs, total RNA was mixed with synthesized cel-miR-238 from plasma denatured specimens. In mixed samples; Cel-Mir-238 was detected. Though, target miR-238 was not visible in RNA of humans as it was not spiked with the miRNA in synthesized form. In addition, there was exceptional linearity amid the qRT-PCR and the cel-mir-238 C values C_t values. Non-specific intensification was noted, whereas only augmentation of the suitable gene therapy corresponding to the particular primer was detected. The findings suggest that qRT reverse transcriptase analysis is relatively accurate and can distinguish between miRNAs from the identical gene family. Consequently, miRNAs and plasma can be accurately detected.

The MiRNA panel has been optimized as a plasma biomarker for NSCLC: The 4 miRNAs used in mixture had high analytical sensitivity in last phases (II-IV) compared to NSCLC in stage I (P less than 0.05) (Table 5). Though, combination of the 4 genes resulted in a sensitivity of 73.33% and a sensitivity of 96.55%. Differentiating NSCLC patients from healthy. Moreover, the the genes diagnostic sensitivity used collectively was higher significantly than that of AC (91.67%) and SCC (82.35%) (P less than 0.05). Lastly, no connotation with smoking-related changes by age, gender, ethnicity among participants was noted.

Table V Shows the Diagnostic Accuracy of MiRNAs among NSCLC Patients

	Sensitivity, %	Specificity, %
All cases	86.89 (53/61)	96.67 (29/30)
Various histological types		
Squamous cell carcinoma	79.41 (27/34)	96.67 (29/30)
Adenocarcinoma	92.59 (25/27)	96.67 (29/30)
Cases with different stages		
I	71.43 (10/14)	96.67 (29/30)
II	88.24 (15/17)	96.67 (29/30)
III	85.71 (12/14)	96.67 (29/30)
IV	87.50 (14/16)	96.67 (29/30)

DISCUSSION

In this analysis, we authenticated formerly recognized NSCLC by means of miRNAs associated with independent tumor tissues by qRT-PCR analysis. We also show that miRNAs were stable and easily measured in plasma¹³. Finally, we showed that the plasma miRNA panel offers probable biomarker that can distinguish healthy individuals from NSCLC cases. Elevation of MiR-21 observed in many

human neoplasms¹⁴. Mir-21 has recently been classified as Oncomir and high levels may result in tumor progression and development. In addition, measuring plasma expression of miR-21 may be beneficial in the analysis of pancreatic cancer¹⁵. Moreover, high serum levels of miRNA-21 may be a predictor of the docetaxel-based chemotherapy efficacy in patients with metastatic hormonal malignancy¹⁶. Furthermore, much upsurge relationship in the level of miRNA expression was found among six miRNAs panel, counting miR-21, amid paired sera and breast tumors¹⁷⁻¹⁸. Moreover, miR-21 was suggestively excessive in normal controls than in cancer patients. Mir-21 may be the blood-derived biomarkers for the analysis, classification and diagnosis of breast cancer¹⁹. Our research data suggests that miR-21 plasma may assist as a lung cancer biomarker for its diagnosis. Mir-486-5p is rare in 8 forms of human cancers, counting lung cancer. The opinion of plasma testing is in line with previous findings and also proposes that miR-486-5p may be a plasma grounded biomarker of lung cancer²⁰. Mir-126 has been shown in human carcinogenesis as a tumor suppressor gene, and low expression has been related with breast cancer metastasis. Moreover, it has recently been found that the reduction of miR-126 expression is closely related to the clinopathological features of breast cancer²¹. In addition, in a study of SCC samples among miRNA profile in skin cancer patients in China, Yang et al. Mir-126 has been found to be aregulatory miRNA compared to cancer-free skin tissue. In addition, the increase in miR-126 expressed in gastric cancer cells subdued growth of cell in the G0 / G1 cell cycle phase and in vivo metastasis and carcinogenicity²². In addition, miR-126 may play a role in tumor suppression in oncogenesis efficiency by targeting the adaptive Crk protein. Our present research displays that low plasma expression of NSCLC mir-126 can differentiate patients from controls. Mir-210 can normalize the hypoxic tumor cell response and tumor growth²³. High Mir-210 expression was suggestively related with estrogen and negative lymph nodes as a positive receptor for human breast cancer. Moreover, it may be one of the slowest increases in serum expression of miR-210 in diffuse large B-cell lymphoma affected patients and adenocarcinoma of pancreatic duct. Our current notes suggest that the Mir-210 may be a plasma monitor for NSCLC. Since smoking is the main reason of NSCLC, most molecular genetic changes are associated with previously diagnosed smoking with lung cancer²⁴. The use of changes such as biomarkers can lead to over diagnosis. Interestingly, the plasma representations of the four miRNAs are independent of the smoking packets²⁵. While our results may seem promising, this study has some limitations. Primary, as the study sample of two cohorts was very minor, and the resulting sensitivity was low for the analysis of Stage I NSCLC. There is a need to further expand the MiRNAs into independent and large cohorts' studies.

CONCLUSION

Finally, we showed that the expression of the surgically identified miRNA in cancer tumors was simple and accurate in plasma measurements. More importantly, a plasma miRNA detection panel would be used as a less invasive analytical method for NSCLC, counting long-term AC.

However, the presence of a potential biomarker requires further verification.

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