

Detection of Novel mutation in VANGL1 gene indicating genetic association of Myelomeningocele

UROOJ FATIMA¹, SYED MEESAM IFTIKHAR², SABAHAT GUL³, FARRUKH MUSTAFA MEMON⁴

¹Assistant Professor, Department of Anatomy, Sind Medical College, Jinnah Sindh Medical University

²Professor, Department of Anatomy, Jinnah Medical and Dental College

³Associate Professor, Department of Anatomy, Quaid e Azam Medical College, Bahawalpur

⁴Professor, Department of Anatomy, Dow International Medical College

Correspondence to Dr. Urooj Fatima, Email id: urooj.fatima@jsmu.edu.pk, Cell# 03218778293

ABSTRACT

Aim: To detect the novel mutation in VANGL1 gene indicating genetic association of Myelomeningocele.

Methodology: The study design was cross sectional. It comprises of sixty individuals, of them fifty were diagnosed cases of myelomeningocele and ten were healthy individuals taken as controls. The cases were collected from Jinnah Postgraduate Medical Center. The study was carried out in Dow Diagnostic and Research Laboratory (D.D.R.L.). Most of the patients were less than one year of age. The cases were evaluated for various other parameters like site and size of cyst and associated features like presence of hydrocephalus in the individuals. Since folic acid deficiency is the key component in the causation of the disease so mothers were also asked about the consumption of folic acid. Blood was drawn from patients after a written permission from the parents of the concerned patient. It was followed by the conduction of PCR to seek for any mutation in VANGL1 gene.

Results: We found a rare mutation in VANGL1 gene revealing substitution of valine to serine at position 239 i.e. V239S. Hydrocephalus being the associated anomaly was present in 32% of the patients. Most of the affected individuals were males. 98% mothers of the sufferers did not take folic acid during pregnancy. In most of the patients, lump was present on the lumbar region.

Conclusion: Myelomeningocele is a congenital birth defect with lifelong complications. Its prevalence can be decreased by taking certain measures. Periconceptional intake of folic acid has been established to lessen the risk of the disease. We identified a rare mutation in VANGL1 gene that may result in the causation of myelomeningocele.

Keywords: Neural tube defects, mutation, myelomeningocele.

INTRODUCTION

Neural tube is the embryonic precursor of brain and spinal cord. Failure of its closure will lead to neural tube defects (NTDs). The second most common anomaly after congenital heart defects are the lesions in the neural tube¹. Recent studies show that the birth prevalence for NTDs ranges between 0.7 and 2.2 per 1000 newborns globally². Anencephaly and myelomeningocele (MMC) are the most common anomalies of neural tube³. The defects which influence the brain are fatal, whereas MMC, although compatible with life but mostly lead to critical disability. Closure of neural tube occurs through a process known as neurulation, in which the neural plate arch upward and finally unites to form a hollow tube that will form the future brain and spinal cord. The dynamic force for the closure of neural tube is contributed and sustained by the cells which undergo convergence and extension⁴. The process of neurulation is divided among mammals into primary and secondary neurulation⁵. During primary neurulation, the union occurs at spine and end up in final closure at the posterior neuropore. Closure begins at hindbrain/ cervical boundary (Closure 1) and extends bidirectionally into the hindbrain and along the spinal section. Independent closure commencement areas happened at midbrain-forebrain boundary (Closure 2) and at the rostral end of forebrain (Closure 3)⁶. If Closure 1 fails to occur it would result in craniorachis, whereas if Closure 1 is

accomplished but the cranial neural tube is defective it would lead to anencephaly¹. Closure 3 failure is not common but if exist would yield split face with anencephaly. In the spinal region, failure of final closure at posterior neuropore yields open spina bifida, in which upper limit can be of varying axial level. By contrast, defective secondary neurulation leads to closed form of Spina Bifida⁷.

The most common NTD linked with survival is Myelomeningocele (MMC)⁸. It is a lethal form of spina bifida that occurs due to failure of neural tube closure at the caudal end leading to protrusion of meninges and spinal cord⁹. Majority of babies born with MMC remain alive but have lifelong disabilities. Deficiency of maternal folic acid levels has been proved to be a significant factor in the causation of NTD¹⁰. Both the genetic and environmental factors are involved in the etiology of the disease. PCP genes are related to noncanonical Wnt pathway that was initially recognized through studies of fruit fly eye and wing pattern genetic screens and was subsequently found to play significant roles during closure of neural tube in mice¹¹. PCP signaling controls process of convergent extension and cell polarity¹² which is essential for gastrulation, neurulation, motile cilia orientation, left-right asymmetry initiation and other procedures in the growth of kidney, liver and other organ. There are various genes linked to PCP which if mutated would cause NTDs¹³. These genes comprise of VANGL1, VANGL2, CELSR1, FZD and DVL. VANGL1 and VANGL2 are mammalian homologues of Drosophila gene Van Gogh (Vang). In humans, VANGL1 gene is situated on chromosome 1p11- p13.1. It encodes

Received on 03-03-2021

Accepted on 25-07-2021

524 amino acid and its size is 56kb. VANGL2 is situated on chromosome 1q22-q23 and encodes 521 amino acid; its size is 28 kg¹⁴. Expression of VANGL1 gene is limited to midline floor plate cells and to notochord¹⁵. Mutation in VANGL1 gene is associated with NTDs in humans¹⁶.

In this study we have hypothesized that mutation in VANGL1 gene would lead to Myelomeningocele and these mutations are present in our population.

MATERIAL AND METHODS

This is a cross sectional study which consist of fifty diagnosed cases of myelomeningocele and ten healthy individuals taken as controls. The patients were recruited at Jinnah Postgraduate Medical Center (J.P.M.C.) and the study was conducted at Dow University of Health Sciences (D.U.H.S.) Most of the mothers did not take folic acid supplementation periconceptionally. Few patients have positive family history. The experimental protocol was approved by the I.R.B. of D.U.H.S. and a consent form was signed by the patient's attendants.

Inclusion Criteria:

- 1) The age of the patient ranges from 0 -10 years.
- 2) The patients were selected regardless of sex, socioeconomic and ethnic background.

Exclusion Criteria

- 1) Other cases of neural tube defects.
- 2) Patients older than the given range of the study.

A trained phlebotomist was hired who drew 2c.c of blood and then placed in EDTA tubes. DNA was obtained from whole blood by QiAmp DNA mini kit. Extraction was done according to specifications provided by the fabricator. It was followed by the PCR amplification of VANGL1 gene. PCR was performed in a tube which contains a reaction mixture of 20µl. The mixture was prepared by following constituents 500 µM of four deoxynucleotides, 10 pmol of each forward and reverse primers for VANGL1 gene, 2 U of Taq polymerase (Promega), 10 x PCR buffer and 1.5 mM MgCl₂. The thermal cycler was designed to initially incubate the product for 10min for 95°C followed by 35 cycles comprising of 94°C for 30s, 64°C for 1min and 72°C for 1min with final extension for 10min at 72°C for 1min with final extension for 10min at 72°C. The amplified products were run on 2% Agarose and visualized under transilluminator. The length of the product was calculated according to migration pattern of a 50bp DNA ladder. The image of the products was taken using DOC gel documentation system. The amplified products were of 404bp and were then sent for commercial sequencing. Mutation was analyzed by aligning the sequence with the reference sequence.

RESULTS

The selected patients were known cases of myelomeningocele. The diagnostic factor was the presence of lump on the vertebral column which is mostly on the lumbosacral region. Most of the mothers (98%) did not take folic acid periconceptionally which may be the causative factor of MMC. Patients were assessed for any associated anomaly like hydrocephalus and it was found that 32% of patients were associated with it. Molecular analysis was further carried out to seek for any mutation in VANGL1

gene. Sixty blood samples were taken, of them, fifty were known cases of MMC and ten were healthy individuals which were labeled as controls. Extraction of DNA and PCR amplification of VANGL1 gene was carried out and the products were sent for commercial sequencing. One of the sample, among fifty cases indicated mutation in the targeted region of exon 4 of VANGL1 gene and amplification was seen at 404 base pairs. The mutation V239S was evident on the position 239 of exon 4 of VANGL1 gene and it was verified by DNA sequencing. Our result revealed the substitution of valine with serine at position 239 in one of the sample. The mutation has not been observed previously and is present in our population as shown in figure 2.

A mutation (V239S) was shown in sample # 16 in which a nine month old boy had myelomeningocele cyst at lumbosacral region (Figure 1). His family didn't show any sign of neural tube defect. His mother did not take folic acid during pregnancy. Due to low socioeconomic status, no antenatal ultrasound was performed to rule out the anomaly in the fetus.

Figure 1(a) A nine month old boy having MMC cyst at lumbosacral region



Figure 2: The analysis of VANGL 1 exon 4 DNA sample, Lane 1, 2, 3, 5,6,7, and 8 were negative with the VANGL 1 exon 4 mutation whereas Lane 4 was identified as positive for VANGL exon 4 mutation Lane N is negative control, Lane P is positive control and Lane M is the DNA ladder of 50 bp.



(b) MRI of the same boy having MMC cyst

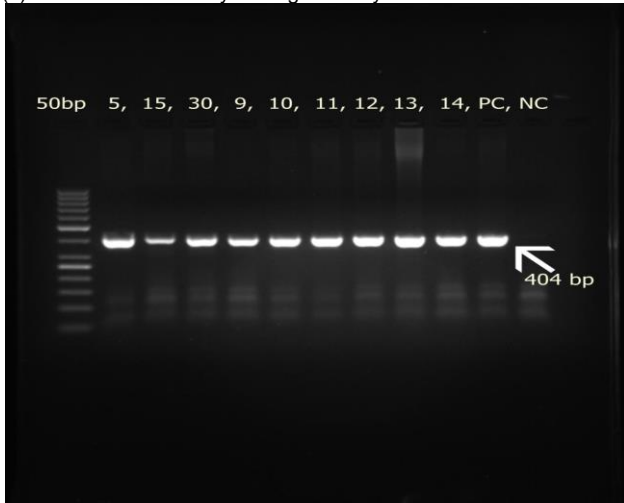


Figure 3: Chromatogram of patient with MMC showing substitution of valine with serine at position 239 in sample number 16 compared with control

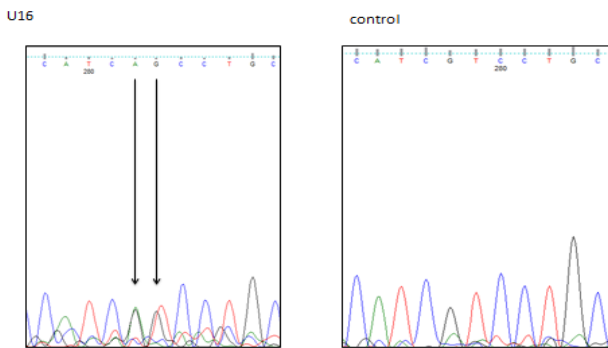
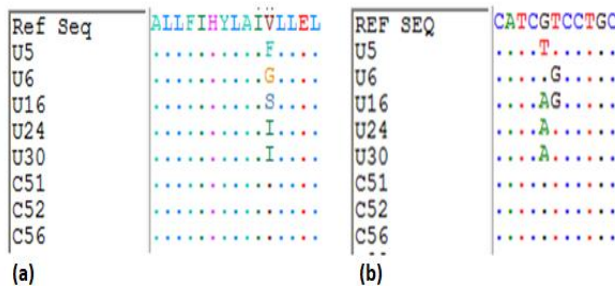


Figure 4: Sequence Alignment showing substitution of Valine to Serine confirmed by DNA sequencing



DISCUSSION

Neural tube defects are a group of birth defects which are related to morbidity, mortality and lifelong disability. It can be prevented by periconceptional intake of folic acid¹⁷. Myelomeningocele being the most frequent type of NTD, have an incidence rate of 0.20–0.40/1000 births/year in Canada and the USA¹⁸. Patients suffering from spina bifida have ten times death risk compared with the rest of the population¹⁹. Infants having open NTDs have 88-96%

chance of 1 year survival rate. In addition to it, such individuals have long lasting complications due to the open defects²⁰.The pathologies associated with the disease require constant medical and rehabilitative care.

Recognition of causative element is established by the fact that most of these defects occur by the union of environmental and genetic factors. To date about 300 genetic mutants linked with NTDs have been established in mice²¹. Such genes exist in various pathways which include planar cell polarity pathway, MAPK pathway, folate metabolic pathway and hedgehog signal transduction pathway²².

In our study we found that among the fifty cases, only one patient showed the positive family history. Dupépe et al stated that there is relatively strong association of family history in neural tube defects²³. It is quite possible that due to small sample size our results differ from them. Nowadays it is easy to diagnose MMC due to progress in the field of sonography. In our case, only six mothers were aware of having such babies prenatally. Requeijo et al in 2016 agreed the reality of ultrasound in visualizing the neural tube defects²⁴. Unsinn et al in 2016 acknowledged the approach of ultrasound in revealing myelomeningocele²⁵. The prevalence of myelomeningocele has been deteriorated worldwide due to prenatal diagnosis through ultrasound. The parents are guided regarding the pros and cons of the disease and make them ready for therapeutic abortion. Unluckily, due to inadequate awareness there is increase in the incidence of myelomeningocele.

We have found mutation in VANGL1 gene V239S in which valine is substituted to serine at position 239. Several gene mutations have been associated with myelomeningocele. Hilman et al identified novel candidate risk gene in patients of myelomeningocele within glucose homeostasis / oxidative stress and folate/ one carbon metabolism networks²⁶. Lei et al studied the CELSR1 coding region sequence among patients of spina bifida and found a novel mutation in CELSR1 gene in patients of Spina Bifida²⁷. Bosoi established a rare mutation in PRICKLE1 in human neural tube defects²⁸. Kibar et al identified five novel missense variants in VANGL1, p.Ser83Leu, p.Phe153Ser, p. Arg181Gln, p.Leu202Phe and p.Ala404Ser occurring in sporadic and familial cases of spinal dysraphisms²⁹. Connealy et al established the genetic variation in GLUT3 gene in patients of myelomeningocele³⁰. Merello et al found identified three heterozygous missense variants in VANGL1, p.Ala187Val, p.Asp389His, and p.Arg517His in individuals suffering from neural tube defects³¹.

CONCLUSION

Myelomeningocele is a complex birth defect. With this research we eventually establish the fact that further exploration and diagnosis of the disorder should be accomplished. In other circumstances, despite of the fact that history manifested myelomeningocele but mutation was not recognized. So keeping this point in consideration, other gene mutation should be done and more research of same area based should be carried out.

Conflict of interest: None

REFERENCES

1. Wu G, Huang X, Hua Y, Mu D. Roles of planar cell polarity pathways in the development of neural tube defects. *J. Biomed. Sci.* 2011 Dec 1;18(1):66.
2. Tian T, Lei Y, Chen Y, Guo Y, Jin L, Finnell RH, Wang L, Ren A. Rare copy number variations of planar cell polarity genes are associated with human neural tube defects. *Neurogenetics.* 2020 May 9.
3. Wang M, de Marco P, Capra V, Kibar Z. Update on the Role of the Non-Canonical Wnt/Planar Cell Polarity Pathway in Neural Tube Defects. *Cells.* 2019 Oct;8(10):1198.
4. Davidson LA, Keller RE. Neural tube closure in *Xenopus laevis* involves medial migration, directed protrusive activity, cell intercalation and convergent extension. *Development.* 1999 Oct 15;126(20):4547-56.
5. Fedorova V, Vanova T, Elrefae L, Pospisil J, Petrasova M, Kolajova V, Hudacova Z, Baniarova J, Barak M, Peskova L, Barta T. Differentiation of neural rosettes from human pluripotent stem cells in vitro is sequentially regulated on a molecular level and accomplished by the mechanism reminiscent of secondary neurulation. *Stem Cell Res.* 2019 Oct 1;40:101563.
6. Greene ND, Copp AJ. Neural tube defects. *Annu. Rev. Neurosci.* 2014 Jul 8;37:221-42.
7. Mohd-Zin SW, Marwan AI, Abou Chaar MK, Ahmad-Annur A, Abdul-Aziz NM. Spina bifida: pathogenesis, mechanisms, and genes in mice and humans. *Scientifica.* 2017 Jan 1;2017.
8. Shah RH, Northrup H, Hixson JE, Morrison AC, Au KS. Genetic association of the glycine cleavage system genes and myelomeningocele. *Birth Defects Res A Clin Mol Teratol.* 2016 Oct;106(10):847-53.
9. Houtrow AJ, Thom EA, Fletcher JM, Burrows PK, Adzick NS, Thomas NH, Brock JW, Cooper T, Lee H, Bilaniuk L, Glenn OA. Prenatal Repair of Myelomeningocele and School-age Functional Outcomes. *Pediatrics.* 2020 Feb 1;145(2).
10. Pei L, Wu J, Li J, Mi X, Zhang X, Li Z, Zhang Y. Effect of periconceptional folic acid supplementation on the risk of neural tube defects associated with a previous spontaneous abortion or maternal first-trimester fever. *Hum. Reprod.* 2019 Aug 1;34(8):1587-94.
11. Wang L, Xiao Y, Tian T, Jin L, Lei Y, Finnell RH, Ren A. Digenic variants of planar cell polarity genes in human neural tube defect patients. *Mol. Genet. Metab.* 2018 May 1;124(1):94-100.
12. Henderson DJ, Long DA, Dean CH. Planar cell polarity in organ formation. *Curr Opin. Cell Biol.* 2018 Dec 1;55:96-103.
13. Tian T, Lei Y, Chen Y, Karki M, Jin L, Finnell RH, Wang L, Ren A. Somatic mutations in planar cell polarity genes in neural tissue from human fetuses with neural tube defects. *Hum. Genet.* 2020 Apr 30.
14. Zhang R, Fang Y, Wu B, Chemban M, Laakhey N, Cai C, Shi O. Gene-gene interaction between VANGL1, FZD3, and FZD6 correlated with neural tube defects in Han population of Northern China. *Genet Mol Res.* 2016 Jan 1;15(3):10-4238.
15. Belotti E, Puvirajesinghe TM, Audebert S, Baudelet E, Camoin L, Pierres M, Lasvaux L, Ferracci G, Montcouquiol M, Borg JP. Molecular characterisation of endogenous Vangl2/Vangl1 heteromeric protein complexes. *PLoS One.* 2012 Sep 28;7(9):e46213.
16. Kibar Z, Bosoi CM, Kooistra M, Salem S, Finnell RH, De Marco P, Merello E, Bassuk AG, Capra V, Gros P. Novel mutations in VANGL1 in neural tube defects. *Hum. Mutat.* 2009 Jul;30(7):E706-15.
17. de la Fournière B, Dhombres F, Maurice P, de Foucaud S, Lallemand P, Zérah M, Guilbaud L, Jouannic JM. Prevention of Neural Tube Defects by Folic Acid Supplementation: A National Population-Based Study. *Nutrients.* 2020 Oct;12(10):3170.
18. North T, Cheong A, Steinbok P, Radic JA. Trends in incidence and long-term outcomes of myelomeningocele in British Columbia. *Childs Nerv Syst.* 2018 Apr;34(4):717-24.
19. Heuer GG, Moldenhauer JS, Adzick NS. Prenatal surgery for myelomeningocele: review of the literature and future directions. *Childs Nerv Syst.* 2017 Jul 1;33(7):1149-55.
20. Bakketun T, Gilhus NE, Rekand T. Myelomeningocele: need for long-time complex follow-up—an observational study. *Scoliosis Spinal Disord.* 2019 Dec;14(1):1-5.
21. Leduc RY, Singh P, McDermid HE. Genetic backgrounds and modifier genes of ntd mouse models: An opportunity for greater understanding of the multifactorial etiology of neural tube defects. *Birth Defects Res.* 2017 Jan 30;109(2):140-52.
22. Wilde JJ, Petersen JR, Niswander L. Genetic, epigenetic, and environmental contributions to neural tube closure. *Annu. Rev. Genet.* 2014 Nov 23;48:583-611.
23. Dupéché EB, Patel DM, Rocque BG, Hopson B, Arynchyna AA, Ralee Bishop E, Blount JP. Surveillance survey of family history in children with neural tube defects. *Journal of Neurosurgery: Pediatrics.* 2017 Jun 1;19(6):690-5.
24. Requeijo MJR, Bunduki V, Francisco RPV, Lopes MAB, Ruano R, Zugaib M. Comparison of two- and three-dimensional ultrasonography in the evaluation of lesion level in fetuses with spina bifida. *Revista Brasileira de Ginecologia e Obstetrícia.* 2016;38(3):120-6.
25. Unsinn KM, Geley T, Freund MC, Gassner I. US of the spinal cord in newborns: spectrum of normal findings, variants, congenital anomalies, and acquired diseases. *Radiographics.* 2000;20(4):923-38.
26. Hillman P, Baker C, Hebert L, Brown M, Hixson J, Ashley-Koch A, Morrison AC, Northrup H, Au KS. Identification of novel candidate risk genes for myelomeningocele within the glucose homeostasis/oxidative stress and folate/one-carbon metabolism networks. *Mol. Genet. Genomic Med.* 2020 Nov;8(11):e1495.
27. Lei Y, Zhu H, Yang W, Ross ME, Shaw GM, Finnell RH. Identification of novel CELSR1 mutations in spina bifida. *PloS one.* 2014 Mar 14;9(3):e92207.
28. Bosoi CM, Capra V, Allache R, Trinh VQ, De Marco P, Merello E, Drapeau P, Bassuk AG, Kibar Z. Identification and characterization of novel rare mutations in the planar cell polarity gene PRICKLE1 in human neural tube defects. *Hum. Mutat.* 2011 Dec;32(12):1371-5.
29. Kibar Z, Bosoi CM, Kooistra M, Salem S, Finnell RH, De Marco P, Merello E, Bassuk AG, Capra V, Gros P. Novel mutations in VANGL1 in neural tube defects. *Hum. Mutat.* 2009 Jul;30(7):E706-15.
30. Connealy BD, Northrup H, Au KS. Genetic variations in the GLUT3 gene associated with myelomeningocele. *Am. J. Obstet Gynecol.* 2014 Sep 1;211(3):305-e1.
31. Merello E, Mascelli S, Raso A, Piatelli G, Consales A, Cama A, Kibar Z, Capra V, Marco PD. Expanding the mutational spectrum associated to neural tube defects: literature revision and description of novel VANGL1 mutations. *Birth Defects Res A Clin Mol Teratol.* 2015 Jan;103(1):51-61.