# **REVIEW ARTICLE**

# A Review on Anti-cancer and Antioxidant Potentials of Plants with Mode of Action

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# ABSTRACT

Cancer is an intensifying health problem, and after myocardial infarction, it is the most common reason for death among people. The current review aimed to highlight the anti-cancer and antioxidant potential of plant-derived natural bio-active compounds. Plants investigated so far with enriched anti-cancer, and antioxidant potential was traced out from previously published literature to present in one document for the ease of the scientific community and researchers working on it. The phytochemicals extracted from herbs have executed numerous healing effect by preventing cancer stimulating hormones and enzymes, exciting DNA reparation mechanisms, endorsing the production of shielding enzymes, persuading antioxidant action, and augmenting immunity, hence presenting the anti-cancer effect. Epidemiological studies suggest that regular intake of anti-cancerous food prevents the onset of many types of cancers. Antioxidant with scavenging properties and free radicals, are predictable to play significant roles in inhibiting ROS induced sicknesses, including cancer, because they can hamper oxidation route by responding with free radicals and chelating free catalytic metals; also by serving as O<sup>2</sup> scavengers. **Keywords:** Anti-cancer plants, Antioxidants, DNA Reparation, Phytochemicals

#### INTRODUCTION

Cancer is an intensifying health problem, and after myocardial infarction, it is the most common reason for death among people<sup>1</sup>. The scientific community has shown interests in pharmacological studies of plant compounds to be used as cancer suppressants<sup>1</sup>. Plant products also perform in synergy with chemotherapeutic drugs by reducing cell resistance to them and cause them to effect efficiently<sup>2</sup>. Cancer could also be prevented by consuming diets that have anti-cancerous properties<sup>3</sup>. Epidemiological studies suggest that regular intake of anti-cancerous food prevents the onset of many types of cancers<sup>4</sup>.

Normally human antioxidant system can rummage these hazardous radicals to keep the balanced oxidation and anti-oxidation. However, excessive contact with cigarette smoke, alcoholic beverages, hazardous radiation, xenobiotics may enhance the production of unnecessary ROS that can disturb the equilibrium of oxidation and anti-oxidation process, resulting chronic and degenerative disorders<sup>5</sup>. To prevent undesirable effects of oxidative stress, boosting of anti-oxidant defensive system of the body is much required with supplementation of exogenous antioxidants<sup>6</sup>. Anti-oxidants are also synthesized in-vivo or administrated orally antioxidants. Plants are potential source of exogenous (i.e., oral administration) antioxidants from a long period.

The current review article was aimed to highlight the anti-cancer and antioxidant potential of plant-derived natural bio-active compounds. Plants investigated so far with enriched anti-cancer and antioxidant potential were traced out from previously published literature to present in one document for the ease of the scientific community and researchers working on it. In this review, edible parts of plants with anti-cancer and antioxidant activity were presented for the information of the community to aware them with the medicinal benefits of commonly available plants.

#### **Mechanism of Antioxidant Action**

Normally, anti-oxidants defense system counter to free radicals prompted oxidative damages<sup>7</sup>. When we look on mood of action, anti-oxidants might be categorized as; free radical terminator, a chelator, or  $O_2$  scavengers<sup>8</sup>. Free radical terminator act as a primary antioxidant reacts with higher energy lipid radicles and converts them into a thermodynamically more stable product. It contributes hydrogen/electron from phenolic hydroxyl groups, which either prevent the initiation or propagation step given in equations below<sup>9</sup>. An anti-oxidant (AH) reacts with radicals formed during autoxidation. The antioxidants free radicle presented in below equations are low energy free radicles which don't allow initiating the chain propagation reactions but entering into a termination reaction with lipid radicals to form lipid anti-oxidant products<sup>8</sup>.

R°+ AH. → RH + A°(Inhibition of the propagation
step)
R° OO° + AH
termination reaction)
$RO^{\circ} + AH \longrightarrow ROH + A^{\circ}(Involvement in termination)$
reaction)
R°+ A° → RA
R°+ A° ───► ROA

- 1. Electron to active-radical leads to cation and anion radical formation, which is flourished by transfer of proton from cation to anion radical.
- 2. H<sup>+</sup> transfers from anti-oxidant to active-radicles
- 3. Deprotonation leads to electron transfer by subsequent anion to active-radicle
- 4. Proton (electron coupled) transfers for phenolic antioxidant

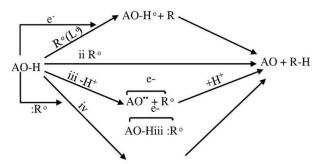


Fig. 1: Process of radical scavenging by an antioxidant<sup>8</sup>.

#### TYPES OF ANTIOXIDANTS Non-enzymatic Antioxidants

Non enzymatic defensive systems alongside oxidative stress comprising on antioxidants; viz,  $\alpha$ -tocopherol, phenolic compounds, carotenoids, glutathione and ascorbic acid<sup>10</sup>, which are liable for scavenging free radicals presence in plants by donating electron/ hydrogen<sup>11</sup>.

#### Ascorbic-acid

Mitochondria are its production house from where it is transported to other constituents of the cell by mechanism of facilitated diffusion<sup>12</sup>. Generally, it exists in condensed form, where its concentrations within cell range from 20 mM and 20–300 mM in cytosol and stroma (chloroplast), respectively<sup>13</sup>. It protects plants alongside ROS produced by photosynthetic and respiratory activities<sup>11</sup>. Physiological processes like growth regulation, plants metabolism and differentiation process is influenced by ascorbic acids.

#### Glutathione

Low molecular weight tri-peptide (yglutamylcysteinylglycine) having non-protein thiols, exists in virtually all components of cell, specifically ETRs (endoplasmic reticulum), mitochondria, chloroplast and chloroplasts. As a major source of thiol, glutathione serves extensive variety of biological and chemical function in living organisms. The neutrophilic properties of thiol are prime group in creation of mercaptide bonding with metals and reaction with some electrophiles<sup>13</sup>. Glutathione is involved in various cellular processes like regulation of enzymatic activity, cell growth/division, cell differentiation, cell death and senescence, detoxification of xenobiotics, regulation of sulfate transport, production of proteins and nucleotides, synthesis of phytochelatins, conjugation of metabolites, and expression of stress-responsive genes<sup>14</sup>.

#### Carotenoids

Carotenoids are natural lipophilic pigments produced from phytoene<sup>15</sup> found in plastids. Carotenoids are also present in microorganisms<sup>14</sup>. More than 600 types of carotenoids exist in nature, with  $\beta$ -carotene the most abundant among others (Olson & Krinsky, 1995). Carotenoids prevent cellular lipid and membrane damage by scavenging peroxyl radicals produced during lipid peroxidation of cell membranes<sup>15-16</sup>. Different studies indicated that carotenoids constrain various kinds' age-related muscular degeneration, cancer types, atherosclerosis, and some other disorders<sup>13</sup>.

#### $\alpha$ -Tocopherols

Tocopherols are a lipid-soluble antioxidant present in almost all plant parts including algae. There are four  $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$  isomers of tocopherols, where  $\alpha$ -tocopherols are the most abundant and active antioxidants in the chloroplast membrane (where there is one molecule of  $\alpha$ -tocopherol for ten molecules of chlorophyll) and protect it against photo-oxidative damage<sup>13</sup>. It works as a well-organized 'chain breaker' while lipid peroxidation process occurring in cell membranes and other lipid particles, consisting of LDL (low density lipoprotein). Maize, wheat germs and cottonseeds and sunflower oil contain higher level of tocopherols and others like as tocols and tocotrienols; these are required to protect seed during germination from lipid peroxidation and for longevity of seed<sup>17</sup>.

#### Phenolic Compounds

A group of secondary metabolites in plants cells, including flavonoids, tannins, hydroxycinnamates esters and lignins. Phenolic acids also include in phenolic compounds antioxidants; it includes the byproducts of cinnamic acid benzoic acid <sup>18</sup>. It is responsible for  $H_2O_2$  scavenging.

Anti-oxidative properties of polyphenols are due to

- i) hydrogen or electron contributors donating high reactivity,
- ii) stabilization and delocalization properties to electron un-pairing
- iii) Chelating ability for transmission metal ion
- iv) Reduction of fluidity of membrane

#### **Enzymatic Anti-oxidants**

Enzymes existent in different sub-cellular components as a part of anti-oxidant machanism including superoxidedismutases, catalases, ascorbate-peroxidase, mono dehydro-ascorbate reductases, dehydro-ascorbate reductases, glutathione-reductases, and guaiacolperoxidases<sup>14</sup>.

#### Superoxide-dismutase (SODs)

Superoxide dismutase is a part of metalloenzymes family and forms frontline defense against reactive oxygen species<sup>14-19</sup>. SODs found in various creatures, like bacteria, yeasts, different animals, and plants. Plants carry numerous genes encoded SODs, which may be accomplished by expansion, and environmental signs<sup>19</sup>. Larson, 1998 reported that activity of superoxide dismutase and catalase have been shown to decline in older leaves of tobacco plants, which indicates signs of membrane damage<sup>20</sup>.

#### Catalase (CAT)

Catalase was ever first characterized antioxidant enzyme<sup>15</sup>. Catalase is tetrameric heme-containing enzyme, present in peroxisomes- a hotspot of  $H_2O_2$  production. It is found in peroxisomes of leaves and eliminates  $H_2O_2$  formed during the process of photo-respiration via alteration of glycolate to glyoxylate<sup>11</sup>. This enzyme involves voguish scavenging of  $H_2O_2$ , which can diffuse from the cytosol. There are three isoforms of catalase, CAT1, CAT-2 and CAT-3 found in mitochondria, chloroplast and cytosol but slight tendency to organic peroxide(s) (R-O-O-R)<sup>14</sup>.

#### Peroxidases

Number of iso-enzymes of peroxidase (POD) plays an important role during normal conditions and in response to environmental stresses. There are four types of peroxidases encoded by the plants present in different cell components, including chloroplast and mitochondria<sup>17</sup>. Ascorbate peroxidase (APX) and guaiacol peroxidase (GPX) are most important peroxidases present in plants defense systems against reactive oxygen species. Ascorbate peroxidase is the essential ingredient of ascorbate glutathione cycle. It executes similar functions as catalase like, scavenging of H<sub>2</sub>O<sub>2</sub>, but at a different location in cytosol and chloroplast<sup>21</sup>.

# MODE OF ACTION AND MOLECULAR TARGETS OF PHYTOCHEMICALS WITH ANTI-CANCER POTENTIALS

Medicinal herbs have been observed to impart therapeutic effect to prevent and treat cancer. Herbal treatment reduces the negative effect, cancer-related problems, and enhances the healing process<sup>22</sup>. Anti- cancerous phytochemicals extracted from plants retard the development of normal cells into the malignant tumor and persuade apoptosis of neoplastic cells along with the prevention of metastasis and angiogenesis<sup>1</sup>.

#### PLANTS WITH ANTI-CANCER PROPERTIES

Infusions made from herbs with greater potential to cure cancer work effectually through several bio-chemical paths and effects, various organ structures and nurture of body by subsidiary system<sup>23</sup>. Various plants have been identified to be beneficial to prevent and treat cancer such as; Aronia melanocarpa, Camellia sinensis, Catharanthus roseus, Datura metel, Momordica charantia, Curcuma longa, Gymnema sylvestre, Stevia rebaudiana, Scutellaria, Nelumbo nucifera, Oroxylum indicum e.t.c<sup>23-24-25-26</sup>. Species of plants with anti-cancer potential are given in Table-I.

Botanical Name	Bio-active Compound	Specific Cancers Suppressed
Aegle marmelos	Skimmianine	Liver cancer (Both in vitro and in vivo) <sup>31</sup>
Allium wallichii	Steroids, terpenoids, flavonoids, reducing sugars and glycosides	Prostate cancer, breast cancer, cervical cancer (In vitro) <sup>25</sup>
Aloe vera	Alexin B, emodin	Leukemia, stomach cancer (In vivo) <sup>32</sup>
Amoora rohituka	Amooranin	Lymphocytic leukemia (In vitro)
Annona crassiflora	Caffeic acid, sinapic acid, rutin	Glioma, renal, ovary cancer (In vitro) <sup>33</sup>
Argemone gracilenta	Argemonine and berberine	B-cell lymphoma, leukemia (In vitro) <sup>34</sup>
Artemisia annua	Artemisinin	Liver, breast and pancreatic cancer (Both in vitro and in vivo) <sup>35</sup>
Bleekeria vitensis	Elliptinium	Myelogenous leukemia and breast cancer (In vivo) <sup>36</sup>
Boerrhavia diffusa	Punarnavine	Malignant melanoma cancer (In vitro) <sup>37</sup>
Broussonetia papyrifera	2S-abyssinone II verubulin	Glioblastoma and brain cancer (In vitro) <sup>38</sup>
Capsicum annuum	Luteolin	Colorectal cancer (Both in vitro and in vivo) <sup>39</sup>
Camelia sinesis	Epicatechingallate, picatechin,	Picatechin, epigallocatechin
	epigallocatechin, Theabrownin	Lung, bladder, skin, prostate and breast cancer (Both in vitro and in vivo)27
Centella asiatica	Asiatic acid	Melanoma, glioblastoma and breast cancer (In vivo) <sup>39</sup>
Cicer arietinum	Bowman-Birk-type protease	Breast and prostate cancer (In vitro) <sup>28</sup>
Colchicum autumnale	Colchicine	Hodgkin's lymphoma, chronic granulocytic leukemia (Both in vitro and in vivo) <sup>40</sup>
Combretum caffrum	Combretastatins	Colon, and leukemia and lung cancer (In vivo) <sup>35</sup>
Crocus sativus	Crocetin	Hippocampal cell death and lung cancer (In vivo) <sup>41</sup>
Curcuma longa	Curcumin, ascorbic acid	Leukemia, glioblastoma, Colon adenocarcinoma, prostate esophagus, liver and skin cancer (In vitro) <sup>30</sup>
Debregeasia saeneb	Tannins	Internal tumors (In vitro) <sup>10</sup>
Dioscorea colletti	Dioscin	Liver and human gastric cancer (In vitro) <sup>24</sup>
Elusine coracana	Ragi bifunctional inhibitor	Myeloid leukemia cell and K562 cell line (Both in vitro and in vivo)42
Ginkgo biloba	Ginkgetin, ginkgolide A &B	Hepatocarcinoma, ovary, prostate, colon and liver cancer <sup>43</sup>
Glycyrrhiza glabra	Licochalcone-A, licoagrochalcone	Prostate, breast, lung, stomach and kidney cancer (In vivo) <sup>29</sup>
Gossypium hirsutum	Gossypol	Mice xenograft (HT-29) and colorectal cancer (Both in vitro and in vivo) <sup>13</sup>
Herba epimedii	Icariin, icaritin, icariside II	Prostate, lung, kidney and gastric cancer (Both in vitro and in vivo) <sup>24</sup>
Morinda citrifolia	Damnacanthal	Lung cancer, sarcomas (In vitro)
Nigella sativa	Thymoquinone	Colon, prostate, breast and pancreas cancer <sup>07</sup>
Ocimum sanctum	Eugenol, orientin, vicenin	Breast, liver and fibrosarcoma cancer (In vitro)44
Oldenlandia diffusa	Ursolic acid	Lungs, ovary, uterus, stomach, liver, colon, rectum and brain cancer (Both in vitro and in vivo) <sup>31</sup>
Panax ginseng	Panaxadiol, panaxatriol	Human colon cancer (Both in vitro and in vivo), Breast, ovary, lung and prostate (In vitro) <sup>29</sup>
Panax pseudoginseng	Panaxadiol	Human colon cancer (Both in vitro and in vivo) <sup>29</sup>
Passiflora caerulea	Chrysin	Colorectal cancer (in vitro) <sup>08</sup>
Peganum harmala	Harmine	Breast cancer (Both in vitro and in vivo) <sup>24</sup>
Peltophorum dubium	Peltophorum dubium trypsin inhibitor	Rat lymphoma cells, human leukemia cells <sup>02</sup>

Table-I: Anti-cancer potential of plant species with bioactive compound and site of action

Plumbago zeylanica	Plumbagin	Liver, fibrosarcoma, leukemia and breast cancer (In vitro) <sup>29</sup>
Polygonum cuspidatum	Resveratrol	Colorectal, skin and liver cancer (In vitro)42
Psoralea corylifolia	Psoralidin, Bavachanin, corylfolinin, psoralen	Stomach, prostate cancer, Lung, osteosarcoma, fibrosarcoma and liver cancer (In vitro) <sup>11</sup>
Scrophularia Atropatana	dichloromethane extract of S. atropatana	breast cancer <sup>21</sup>
Solanum nigrum	Solamargine, solasonine	Breast, liver, lung and skin cancer (In vitro) <sup>45</sup>
Sylibum marianum	Silibinin, Silymarin	Lung, liver, skin, colon and prostate cancer, Colorectal cancer (Both in vitro and in vivo) <sup>18</sup>
Tylophora indica	Tylophorine	Breast cancer (In vivo)
Vaccinium macrocarpon	Hydroxycinnamoyl ursolic acid	Cervical, prostate cancer (In vitro) <sup>14</sup>
Vicia faba	Field bean protease inhibitors	Skin cancer (Both in vitro and in vivo) <sup>02</sup>
Vigna unguiculata	Black-eyed-pea trypsin/ Chymotrypsin inhibitor	Human breast cancer (In vitro) <sup>34</sup>
Vitis vinifera	Procyanidins	Human colon cancer (In vitro)
Withania somnifera	Withaferin A, D	Breast, cervix, prostate and colon cancer (In vivo) <sup>29</sup>
Zingiber officinale	6-Shogaol	Ovary cancer (In vitro) <sup>33</sup>
Ziziphus mauritiana	a-linolenic acid, Methyl stearate	Leukemia, human cervical and liver cancer (In vitro) <sup>41</sup>
Ziziphus spina-christi	Doxorubicin, spinanineA, rutnine, quercetin	Lung cancer and breast cancer (In vivo) <sup>19</sup>

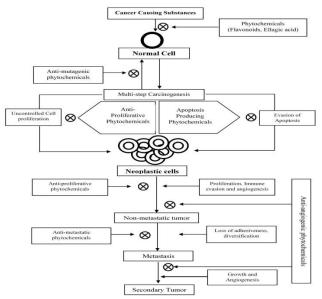


Fig. 2: Mode of action adopted by phytochemicals against cancer  $^{\scriptscriptstyle 12}$ 

Allium wallaichii has moderate antioxidant and cytotoxic properties and includes many phytochemicals such as; steroids, glycosides, flavonoids and terpenoids. Allium wallaichii has the ability to be used as an anti-cancer agent<sup>27</sup>. Artemisia annua, also known as qinghao in China, where it has long been informed<sup>28</sup>. Artemisinin (ARS), is the active ingredient which is anti-cancerous against pancreatic cancer, liver cancer and, breast cancer.

Kumari<sup>27</sup> studied that green tea is obtained from new and soft leaves of Camelia sinesis. Study on Curcuma longa L has exposed for curcumin substantial potential against a varied type of both diseases (malignant and non malignant)<sup>30</sup>. Debregeasia saeneb suppresses internal tumors. Ginkgo biloba, Glycyrrhiza glabra, Peganum harmala L., Scrophularia Atropatana, Solanum nigrum, Vigna unguiculate, Withania somnifera (L.), Zizipus mauritiana Lam, Z. nummularia, Z. mauritiana and Z. jujuba are reported anticancer medicinal plants.

# CONCLUSION

This review article describes the anti-cancer and antioxidant potential of plant-derived natural bio-active compounds. A number of medicinal plants edible parts carry anti-cancer and antioxidant activities; are presented here for information of community to aware them with medicinal benefits of commonly available plants. This article provides natural and plant base antioxidants list with their specific activity and mode of action and detail of bioactive compounds to suggest that the use of synthetic antioxidants for food preservation and medicinal purpose need to be restricted due to its toxic impacts.

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### REFERENCES

- Rahman MM, Khan A (2013) Anti-cancer potential of South Asian plants. Nat Prod. Biopros 3:74–88 Liu, R. H. (2004). Potential Synergy of Phytochemicals in Cancer Prevention: Mechanism of Action. The Journal of Nutrition, 134(12), 3479S–3485S.
- Amin, A. R. M. R., Kucuk, O., Khuri, F. R., and Shin, D. M. (2009). Perspectives for Cancer Prevention with Natural Compounds. Journal of Clinical Oncology, 27(16), 2712– 2725.
- Russo, M., Spagnuolo, C., Tedesco, I., and Russo, G. L. (2010). Phytochemicals in Cancer Prevention and Therapy: Truth or Dare? Toxins, 2(4), 517–551.
- Subhasree, B., Baskar, R., Keerthana, R.L., Susan, R.L., Rajasekaran, P., 2009. Evaluation of antioxidant potential in selected green leafy vegetables. Food Chemistry 115, 1213– 1220.Li, S., Tan, H.Y., Wang, N., Zhang, Z.J., Lao, L., Wong, C.W., Feng, Y., 2015. The role of oxidative stress and antioxidants in liver diseases. Int. J. Mol. Sci. 16, 26087– 26124.
- Kasote, D.M., Hegde, M.V., Katyare, S.S., 2013. Mitochondrial dysfunction in psychiatric and neurological diseases: cause(s), consequence(s), and implications of antioxidant therapy. Biofactors 39, 392-06.
- 6. Daramola, B., 2014. Anthology of historical development and some research progress glimpses on phytochemical

antioxidants phenomenon. International journal of biotechnology and molecular biology research 5 (3), 13-26.

- 7. Aziz, M.A., Diab, A.S., Mohammed, A.A., 2019. Antioxidant Categories and Mode of Action. Antioxidants. doi:10.5772/intechopen.83544.
- 8. Giese, J., 1996. Antioxidants: tools for preventing lipid oxidation. Food technology 50 (11), 73 81.
- Blokhina, O., Virolainen, E., Gagerstedt, K.V., 2003. Antioxidants, oxidative damage and oxygen deprivation stress: a review. Ann Bot (Lond) 91, 179–194.
- Jaleel, A.C., Riadh, K., Gopi, R., Manivannan, P., Ine's, J., Al-Juburi, H.J., et al., 2009. Antioxidant defense responses: physiological plasticity in higher plants under abiotic constraints. Acta Physiol Plant 31, 427–436.
- Ahmad, P., Abdul Jaleel, C., Salem, M.A., Nabi, G., Sharma, S., 2010. Roles of enzymatic and non-enzymatic antioxidants in plants during abiotic stress. Critical Reviews in Biotechnology 30 (3), 161–175.
- 12. Horemans, N., Foyer, C.H., Asard, H., 2000. Transport and action of ascorbate at the plant plasma membrane. Trends Plant Sci. 5, 263–267.
- 13. Das, K., Roychoudhury, A., 2014. Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. Frontiers in environmental science doi: 10.3389/fenvs.2014.0005.
- 14. Stahl, W., Sies, H., 2003. Antioxidant activity of carotenoids. Molecular Aspects of Medicine 24, 345-351.
- Engwa, G.A., 2018. Free Radicals and the Role of Plant Phytochemicals as Antioxidants Against Oxidative Stress-Related Diseases. Phytochemicals - Source of Antioxidants and Role in Disease Prevention. doi:10.5772/intechopen.76719.
- Pandhair, V., Sekhon B.S., 2006. Reactive Oxygen Species and Antioxidants in Plants: An Overview. J. Plant Biochemistry and Biotechnology15, 71-78.
- Manach, C., Scalbert, A., Morand, C., Remesy, C., Jimenez, L., 2004. Polyphenols: Food sources and bioavailability. Am. J. Clin. Nutr. 79, 727–747.
- Caverzan, A., Casassola, A., Patussi Brammer, S., 2016. Reactive Oxygen Species and Antioxidant Enzymes Involved in Plant Tolerance to Stress. Abiotic and Biotic Stress in Plants - Recent Advances and Future Perspectives. doi:10.5772/61368
- 19. Larson, R.A., 1988. The antioxidants of higher plants. Phytochemistry 27 (4), 969-978.
- Kangasjärvi, S., Lepistö, A., Hännikäinen, K., Piippo, M., Luomala, E.M., Aro, E.M., Rintamäki, E., 2008. Diverse roles for chloroplast stromal and thylakoidbound ascorbate peroxidases in plant stress responses. Biochem J. 412, 275– 285.
- 21. Aggarwal, B. B., and Shishodia, S. (2006). Molecular targets of dietary agents for prevention and therapy of cancer. Biochemical Pharmacology, 71(10), 1397–1421.
- Ganguly S (2014) Ayurveda for cancer therapy. World J Pharma Res 3:1476–1479
- Kaushik, R., Narayanan, P., Vasudevan, V., Muthukumaran, G., and Usha, A. (2010). Nutrient composition of cultivated stevia leaves and the influence of polyphenols and plant pigments on sensory and antioxidant properties of leaf extracts. Journal of Food Science and Technology, 47(1), 27–33.
- Yoon, J.-S., Kim, H.-M., Yadunandam, A. K., Kim, N.-H., Jung, H.-A., Choi, J.-S., ... Kim, G.-D. (2013). Neferine isolated from Nelumbo nucifera enhances anti-cancer activities in Hep3B cells: Molecular mechanisms of cell cycle arrest, ER stress induced apoptosis and anti-angiogenic response. Phytomedicine, 20(11), 1013–1022
- Bhandari, J., Muhammad, B., Thapa, P. et al (2017). Study of phytochemical, anti-microbial, antioxidant, and anti-cancer

properties of Allium wallichii . BMC Complement Altern Med 17, 102.

- Mahomoodally, M. F., and Gurib-Fakim, A. (2013). Harnessing Traditional Knowledge to Treat Existing and Emerging Infectious Diseases in Africa. Fighting Multidrug Resistance with Herbal Extracts, Essential Oils and Their Components, 223–235.
- Kumari, M. Pattnaik, B. Rajan, S. Y. Padmavathi B.N., Surendra, S. S. Usmani, S. (2017). EGCG - A Promising Anti-Cancer Phytochemical. Annals of International Medical and Dental Research, Vol (3), Issue (2). 8-10
- Aryal, S., Baniya, M.K., Danekhu, K., Kunwar, P., Gurung, R., Koirala, N., 2019. Total Phenolic Content, Flavonoid Content and Antioxidant Potential of Wild Vegetables from Western Nepal. Plants, 8, 96; doi: 10.3390/plants8040096.
- Zhang, Y-J., Gan, R-Y Li, S., Zhou, Y., Li, A-N., Xu, D-P., Li, H-B., 2015. Antioxidant Phytochemicals for the Prevention and Treatment of Chronic Diseases. Molecules 20, 21138– 21156.
- Ooko, E., Kadioglu, O., Greten, H. J., and Efferth, T. (2017). Pharmacogenomic Characterization and Isobologram Analysis of the Combination of Ascorbic Acid and Curcumin—Two Main Metabolites of Curcuma longa—in Cancer Cells. Frontiers in Pharmacology, 8.
- Mukhija, M., Singh, M. P., Dhar, K. L., Kalia, A. N. (2015). Cytotoxic and antioxidant activity of Zanthoxylum alatum stem bark and its flavonoid constituents. Journal of Pharmacognosy and Phytochemistry; 4(4): 86-92.
- Shalabi, M., Khilo, K., Zakaria, M. M., Elsebaei, M. G., Abdo, W., and Awadin, W. (2015). Anti-cancer activity of Aloe vera and Calligonum comosum extracts separetely on hepatocellular carcinoma cells. Asian Pacific Journal of Tropical Biomedicine, 5(5), 375–381.
- Leyva-Peralta, M.A., Robles-Zepeda, R.E., Garibay-Escobar, A. et al. In vitro anti-proliferative activity of Argemone gracilenta and identification of some active components. BMC Complement Altern Med 15, 13 (2015).
- Efferth, T. (2017). From ancient herb to modern drug: Artemisia annua and artemisinin for cancer therapy. Seminars in Cancer Biology, 46, 65–83.
- 35. Lauritano, C., Andersen, J. H., Hansen, E., Albrigtsen, M., Escalera, L., Esposito, F., Ianora, A. (2016). Bioactivity Screening of Microalgae for Antioxidant, Anti-Inflammatory, Anticancer, Anti-Diabetes, and Antibacterial Activities. Frontiers in Marine Science, 3.
- Mishra, S., Aeri, V., Gaur, P. K., and Jachak, S. M. (2014). Phytochemical, Therapeutic, and Ethnopharmacological Overview for a Traditionally Important Herb:Boerhavia diffusaLinn. BioMed Research International, 2014, 1–19.
- Pang, S.-Q., Wang, G.-Q., Lin, J., Diao, Y., and Xu, R. (2014). Cytotoxic activity of the alkaloids fromBroussonetia papyriferafruits. Pharmaceutical Biology, 52(10), 1315–1319.
- Osman, N. H. A., Said, U. Z., El-Waseef, A. M., and Ahmed, E. S. A. (2014). Luteolin supplementation adjacent to aspirin treatment reduced dimethylhydrazine-induced experimental colon carcinogenesis in rats. Tumor Biology, 36(2), 1179– 1190.
- Arpita R, Navneeta B. Centella Asiatica: A Pharmaceutically Important Medicinal Plant. Curr Trends Biomedical Eng and Biosci. 2017; 5(3): 555661
- Lin, X., Peng, Z., and Su, C. (2015). Potential Anti-Cancer Activities and Mechanisms of Costunolide and Dehydrocostuslactone. International Journal of Molecular Sciences, 16(12), 10888–10906.
- 41. Hoshyar, R., and Mollaei, H. (2017). A comprehensive review on anti-cancer mechanisms of the main carotenoid of saffron, crocin. Journal of Pharmacy and Pharmacology, 69(11), 1419–1427.

- 42. Srikanth, S., and Chen, Z. (2016). Plant Protease Inhibitors in Therapeutics-Focus on Cancer Therapy. Frontiers in Pharmacology, 7. doi:10.3389/fphar.2016.00470
- Xiong, M., Wang, L., Yu, H.-L., Han, H., Mao, D., Chen, J., Wang, Y.-A. (2015). Ginkgetin exerts growth inhibitory and apoptotic effects on osteosarcoma cells through inhibition of STAT3 and activation of caspase-3/9. Oncology Reports, 35(2), 1034–1040.
- 44. Preethi R and Padma PR: Biosynthesis and Bioactivity of Silver Nanobioconjugates from Grape (Vitis Vinifera) Seeds and its active Component Resveratrol. Int J Pharm Sci Res 2016; 7(10): 4253-62.
- 45. Al Sinani, S.S., Eltayeb, E.A., Coomber, B.L. et al. Solamargine triggers cellular necrosis selectively in different types of human melanoma cancer cells through extrinsic lysosomal mitochondrial death pathway. Cancer Cell Int 16, 11 (2016).