

Interrelationship of Circulating Biochemical Markers and Response of Lutein in Rats Receiving Carbon Tetrachloride (CCl₄) to Induce Hepatic Injury

ABDUL BASIT¹, MUHAMMAD SAEED QURESHI², SHAZIA ASHRAF⁴, SULAYMAN WAQUAR¹, ZOHAIB RANA¹, DANISH MASOOD³, ARIF MALIK¹

ABSTRACT

This study was conducted to evaluate the hepato-protective effects of Lutein and Clavazin in different dose pattern to reduce the oxidative stress against carbon tetrachloride (CCl₄) induced toxicity in rats. Thirty albino Wistar rats were included, divided into five groups and every group had six rats. Group A was control while group B, C, D and E were treated with 4ml/kg CCl₄ for 10 days with body weight of 160g to 200g. Further group C and D were administered with Lutein 200mg/kg and 400mg/kg respectively for 1 month. Infected CCl₄ group E was treated with a synthetic drug (Clavazin 1Tab/kg). The elevation of ALT, AST, ALP and MDA level in group B while reduced the levels of TP, SOD, CAT and GSH were recorded due to CCl₄ administration. Therapy with Lutein and synthetic drug (Clavazin) in different dose pattern for 1 month significantly decreases the levels of ALT, AST, ALP and MDA. On the other hand, increased levels of TP and anti-oxidants such as GSH, CAT and SOD were observed. The present study demonstrated that Lutein and Clavazin have potent defensive ability against CCl₄ induced liver toxicity in rats by their anti-oxidative and anti-inflammatory activity and may have protective effect on human liver.

Keywords: Carbon Tetrachloride (CCl₄), Sodium oxide dismutase (SOD), Catalase (CAT), Glutathione (GSH), Total protein (TP), Malondialdehyde (MDA), Clavazin, Lutein.

INTRODUCTION

In the recent years, most of halogenated alkane compounds have been banned due to their toxicity such as carbon tetrachloride (CCl₄), chloroform (CHCl₃) or iodoform (CHI₃). CCl₄ induces their effects on different organs such as liver, heart, brain, kidney, lungs and testes (Weber *et al.*, 2003). Carbon tetrachloride (CCl₄) proved to be a lethal liver toxin resulting in production of free radicals that cause oxidative stress leading to centrilobular liver necrosis (Chen *et al.*, 2013). CCl₄ mediated cell damage by covalent binding of reactive intermediates to cellular components which initiate the lipid per-oxidation, leading to apoptosis (Boll *et al.*, 2001). CCl₄ converts into CCl₃ by cytochrome P450, reacts with high partial pressure of oxygen and converts into trichloromethyl peroxy radical (Cl₃COO[•]) (Weber *et al.*, 2003). Liver is the main organ for nourishment, metabolism and detoxifying external chemical mediators as it is the center of most important activities like Cytochrome P450 generation that suppress due to CCl₄ toxicity (Sagor *et al.*, 2015).

Antioxidants play crucial role in protecting the cells against inflammation caused by oxidative damage or free radicals which generate protective response to cellular injury, eliminating the injured tissue and elevating the tissue repairing. Anti-oxidative effects of several biomedical and herbal chemicals have been observed to work against the hepato-toxic effects (Chen *et al.*, 2013). Lutein belongs to the carotenoid family, present in dark green leafy plants like kale and spinach and recommended daily intake is 1.7 mg (Koushan *et al.*, 2013). The transport mechanism of lutein from intestine to tissue is not clearly understood. The process of carotenoids transport in blood and distributed to different tissue is done by the help of low density lipoprotein (LDL). 67% of beta carotenoids are distributed by LDL and 53% of lutein was present in HDL for transportation. The present study was designed to assess the interrelationship of circulating biochemical markers and response of Lutein and Clavazin in rats receiving carbon tetrachloride (CCl₄) to induce hepatic injury.

MATERIALS AND METHODS

Adult Wistar male albino rats were taken from the National Institute of Health, Islamabad, Pakistan and its ranging from 160 to 200g. The study was

¹Institute of Molecular Biology & Biotechnology, University of Lahore

²Department of Biochemistry, Azra Naheed Medical College, Superior University-Lahore

³Allama Iqbal Medical College-Lahore

⁴Histopathology Department, Sheikh Zayed Hospital-Lahore

Correspondence to Dr. Arif Malik, Email: arifuaf@yahoo.com, Cell: 03218448196

approved by the Ethics Committee for the Scientific Research at the University of Lahore.

Plant extract: The standardized extract of Lutein was purchased from the Sigma-Aldrich Corporation (St. Louis, MO, USA).

Experimental design: Thirty (30) albino rats divided into five groups. Every group had six rats. For fourteen days every group administered with CCl_4 to induce toxicity. To check the hepatoprotective effects give lutein to group three and group four for one month in different doses pattern. Group five treated with synthetic drug Clavazin 1tab/kg that contains several extracts such as *Silybum marianum* (200mg), *Picrorhiza kurroa* (50mg), *Glycyrrhiza glabra* (50mg) and *Cichorium intybus* (75mg).

Group 1	Control rats, which received normal diet.
Group 2	Received CCl_4 4ml/ kg.B.wt
Group 3	Received CCl_4 + Lutein 200mg/kg.B.wt
Group 4	Received CCl_4 + Lutein 400mg/kg.B.wt
Group 5	Received CCl_4 + Clavazin 1tab/ kg.B.wt

Blood and serum separation: During the dissection 5mL blood was collected from the rat heart and serum was separated by centrifugation at 1500 rpm for 10 min. After the separation of serum it was stored at -60°C until further biochemical analysis.

Biochemical assays: The aspartate amino-transferase (AST), alanine amino-transferase (ALT), alkaline phosphatase (ALP) and total protein (TP) were determined by using commercial kits (Biomerieux, USA). The levels of GSH, CAT, SOD and MDA were measured by Moron *et al.*, (1979), Aebi, (1974), Kakkar *et al.*, (1984), Ohkawa *et al.*, (1979).

RESULTS

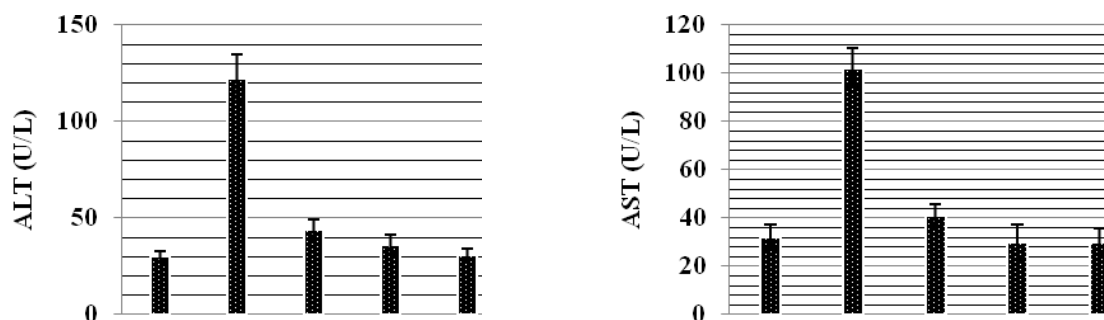
Serum biochemical parameters of ccl_4 induced toxicity in rats: The hepatic metabolic enzyme levels such as ALT, AST and ALP are significantly elevated and total protein (TP) level reduced in CCl_4 treated rats. The TP level reached 2.99IU/L in CCl_4 treated rats as compared to standard value (6.21IU/L), but observed TP level again increased (5.26, 6.25 and 5.21IU/L) in group C (CCl_4 +Lutein 200mg/kg), group D (CCl_4 +Lutein 400mg/kg) and group E (CCl_4 +Lutein Clavazin 1Tab/kg) respectively (Figure-01 A, B, C and D).

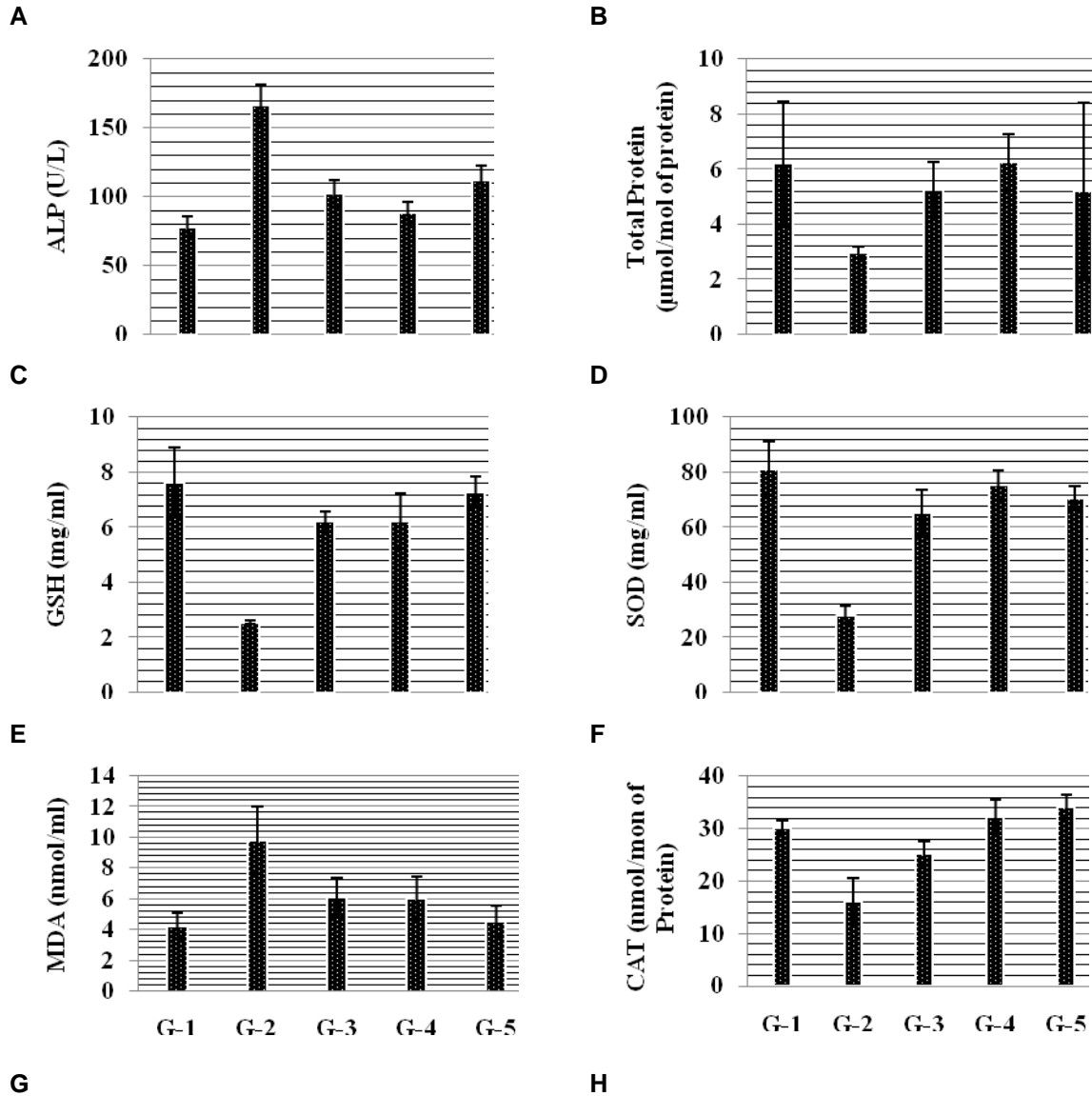
Consequences of lutein on enzymatic antioxidant activities in ccl_4 receiving rats: The antioxidant enzymes such as SOD (28.26mg/ml) and CAT (16.23 $\mu\text{mol/mol}$ of protein) counted low in CCl_4 treated rats as compared to standard value (81.26mg/ml and 30.25 $\mu\text{mol/mol}$ of protein) but on another side, Lutein protects the alteration of SOD and CAT. So, antioxidant activity is enhanced for SOD and CAT in group C (65.26mg/ml, 25.26 \pm 2.35), group D (75.26mg/ml, 32.25 \pm 3.26) and group E (70.26mg/ml, 34.26 \pm 2.25) respectively (Figure-01 F and H).

Consequences of lutein on non-enzymatic antioxidant activity in ccl_4 receiving rats: The non-enzymatic antioxidant GSH activity reduced (7.65 mg/ml to 2.56mg/ml) in CCl_4 treated rats. But Lutein recovered the GSH activity in group C (6.23mg/ml), group D (6.25mg/ml) and group E (7.26mg/ml) respectively (Figure-01 E).

Consequences of lutein on hepatic oxidative stress marker in ccl_4 receiving rats: The lipid peroxidation end product MDA is significantly high (9.823nmol/ml) in CCl_4 treated rats as compared to standard value (4.223nmol/ml). However, Lutein recovered the MDA level and in group C observed the (6.123nmol/ml), in group D (6.023nmol/ml) and in group E (4.526nmol/ml) respectively (Fig.1 G).

Fig. 1: Response of lutein in rats receiving carbon tetrachloride (CCl_4) to induce hepatic injury G-1 (Control), G-2 (CCl_4 4ml/kg b.Wt), G-3 (CCl_4 4ml/kg b.Wt+Lutein @ 200 mg/kg b.Wt), G-4 (CCl_4 4ml/kg b.Wt+Lutein@400mg/kg b.Wt), G-5 (CCl_4 4ml/kg b.Wt+ CLAVAZIN @ 200 mg/kg b.Wt)





DISCUSSION

In the present study, firstly we induce CCl₄ toxicity resulting in relative increase in aspartate amino-transferase (AST), alkaline phosphatase (ALP) and alanine amino-transferase (ALT) and decrease in total protein (TP) in group B. The levels of these biomarkers were reversed, AST and ALP were lowest and TP was highest in rats receiving Lutein at a rate of 400 mg/kg of body weight, ALT was more close to normal in rats administering Clavazin at a rate of 1mg/kg of body weight. The elevated levels of ALT and AST indicate the disturbance in K⁺ and Na⁺ ions that diminish the cell membrane integrity (Naik and Panda, 2008). ALP elevation levels suggest obstruction of liver sinusoids (Giannini *et al.*, 2005). CCl₄ is highly reactive hepatotoxin, induce liver

toxicity (Connor *et al.*, 1986) and its metabolism consists of two phases. In initial phase, CCl₄ converted into CCl₃ by cytochrome p450-dependent monooxygenase in hepatic parenchyma cells while in second phase, CCl₄ activates the Kupffer cells that produce pro-inflammatory mediators (Eidi *et al.*, 2013).

Lutein, a xanthophyll are natural carotenoids having ability to reduce the oxidative stress. Lutein have protective efficacy for lipid membrane against free radicals attack (Sagor *et al.*, 2015). After CCl₄ administration, the Lutein therapy recovers the hepatic damage. During liver damage, increased ROS production is determined by the end product of lipid peroxidation (MDA). The CCl₄ treated group B shows MDA increase but in group C and D Lutein therapy

decreases the oxidative stress, as MDA levels are reduced. Group E that contain synthetic drug (Clavazin) also reduce the MDA levels. The primary antioxidants like GSH, SOD and CAT are also reduced by CCl₄ administration. Clavazin treated rats show highest levels of GSH and CAT while lowest levels of MDA and also represent higher SOD levels only after group D treated with Lutein at a rate of 400 mg/kg of body weight. Kim (Kim *et al.*, 2012) has demonstrated the same positive effects of Lutein on oxidative stress in liver of guinea pigs. Thus, by comparing Lutein and Clavazin, it has been shown that if both are used as a co-treatment it can be more beneficial in neutralizing the deleterious effects produced by CCl₄ toxicity.

CONCLUSION

The present study concludes that CCl₄ administration causes biochemical alterations while Lutein and Clavazin have potent anti-oxidant activity to recover the hepatic damage. Both of Lutein and Clavazin have potential to treat the liver injury in rats but still more research is required to explore their protective effects on human liver that is the major problem of current days.

REFERENCES

1. Aebi H (1974). Catalase in Bergmeyer HU Methods in Enzymatic Analysis: New York, Academic Press. 3:276-286.
2. Boll M, Lutz WD, Becker E, Stampfl A (2001). Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites. *Zeitschrift für Naturforschung*. 56(7-8):649-659.
3. Chen S, Zou L, Li L, Wu T (2013). The protective effect of glycyrrhetic acid on carbon tetrachloride-induced chronic liver fibrosis in mice via upregulation of Nrf2. *PloS One*. 8(1): 536-562.
4. Connor HD, Thurman RG, Galizi MD, Mason RP (1986). The formation of a novel free radical metabolite from CCl₄ in the perfused rat liver and in vivo. *Journal of Biological Chemistry*. 261(10):4542-4548.
5. Eidi A, Mortazavi P, Rohani AH, Safi S, Tehrani ME (2012). Hepatoprotective effects of pantothenic acid on carbon tetrachloride-induced toxicity in rats. 11:748-759.
6. Giannini EG, Testa R, Savarino V (2005). Liver enzyme alteration: a guide for clinicians. *Canadian medical association journal*. 172(3):367-379.
7. Kim JE, Clark RM, Park Y, Lee J, Fernandez ML (2012). Lutein decreases oxidative stress and inflammation in liver and eyes of guinea pigs fed a hypercholesterolemia diet. 6(2):113-119
8. Koushan K, Rusovici R, Li W, Ferguson LR, Chalam KV (2013). The role of lutein in eye-related disease. *Nutrients*. 5(5):1823-1839.
9. Naik SR, Panda VS (2008). Hepatoprotective effect of Ginkgoe select Phytosome in rifampicin induced liver injury in rats: Evidence of antioxidant activity. *Fitoterapia*. 79(6):439-445.
10. Sagor AT, Chowdhury MR, Tabassum N, Hossain H, Rahaman M, Alam A (2015). Supplementation of fresh ucche (*Momordica charantia* L. var. *muricata* Willd) prevented oxidative stress, fibrosis and hepatic damage in CCl₄ treated rats. *BMC complementary and alternative medicine*. 15(1):115.
11. Sujak A, Gabrielska J, Grudziński W, Borc R, Mazurek P, Gruszecki WI (1999). Lutein and zeaxanthin as protectors of lipid membranes against oxidative damage: the structural aspects. *Archives of Biochemistry and Biophysics*. 371(2):301-307.
12. Weber LW, Boll M, Stampfl A (2003). Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. *Critical reviews in toxicology*. 33(2):105-136.
13. Moron MS, Depierre JW, Mannervik B (1979). Levels of glutathione reductase and glutathione S-transferase in rat lung and liver. *Biochem Biophys Acta*. 582: 67-68.
14. Aebi H (1974). Catalase in Bergmeyer HU Methods in Enzymatic Analysis: New York, Academic Press. 3: 276 -286.
15. Kakkar PB, Das P, Viswanathan PN (1984). A modified spectrophotometer assay of superoxide dismutase. *Ind J Biochem Bio*. 21: 130-132.
16. Ohkawa H, Ohishi N, K Yagi K (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *J Anal Biochem*. 95: 351-358.